

CIRCULAR OF INFORMATION

FOR THE USE OF HUMAN BLOOD AND BLOOD COMPONENTS

This *Circular* was prepared jointly by AABB, the American Red Cross, America's Blood Centers, and the Armed Services Blood Program. The Food and Drug Administration recognizes this *Circular of Information* as an acceptable extension of container labels. *Federal Law prohibits dispensing the blood and blood components described in this circular without a prescription.*



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Cryoprecipitated Antihemophilic Factor (AHF) is prepared by thawing Whole Blood derived FFP or Whole Blood derived Plasma Frozen within 24 hours after phlebotomy (PF24) between 1° and 6° C and recovering the precipitate.

Testing of Donor Blood: Blood components collected between December 3, 2018 and June 26, 2021 were tested with a licensed nucleic acid test (NAT) for Zika Virus RNA and found to be nonreactive.

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Notice to All Users

The *Circular of Information for the Use of Human Blood and Blood Components* (hereafter referred to as *Circular*) is an extension of container labels, as the space on those labels is limited.

Blood and blood components are biological products and living human tissue intended for use in patient treatment. Professional judgment based on clinical evaluation determines the selection of components, dosage, rate of administration, and decisions in situations not covered in this general statement.

This *Circular*, as a whole or in part, cannot be considered or interpreted as an expressed or implied warranty of the safety or fitness of the described blood or blood components when used for their intended purpose. Attention to the specific indications for blood components is needed to prevent inappropriate transfusion.

Because of the risks associated with transfusion, physicians or prescribing health care professionals should be familiar with alternatives to transfusion. Blood banks and transfusion services are referred to the AABB *Standards for Blood Banks and Transfusion Services* for additional information and policies, especially in the areas of recipient sample identification, compatibility testing, issue and transfusion of blood and blood components, investigation of transfusion reactions, and proper record-keeping practices. Transfusionists are referred to the AABB *Technical Manual* for applicable chapters on adult and pediatric transfusion.

The specific product manufacturer's instructions for use should be reviewed for information pertaining to the use of transfusion devices (eg, filters, blood administration sets, and blood warmers).

This *Circular* is supplied to conform with applicable federal statutes and regulations of the Food and Drug Administration (FDA), United States Department of Health and Human Services. The blood components in this *Circular* marked with the symbol "Ω" are blood components for which the FDA currently has not received data to demonstrate that they meet prescribed requirements of safety, purity, and potency, and therefore are not licensed for distribution in interstate commerce.

General Information for Whole Blood and All Blood Components

Donors

Blood and blood components described in this *Circular* are collected from blood donors for use in patients (allogeneic transfusions) or from patients donating for themselves (autologous transfusions). Most allogeneic donations are from volunteer blood donors, and the components are labeled "volunteer donor." If donors receive monetary payment for a blood donation, the components must be labeled as "paid donor."

All blood donors satisfactorily complete a health assessment that includes a medical history questionnaire on past and present illnesses. All donors undergo a physical assessment before donation to satisfy minimum physiologic criteria. Allogeneic donors are assessed for risk factors associated with transmissible infectious agents and instructed to call the blood center after donation if they develop illness or have concerns that their blood may not be safe for transfusion.

Autologous donations are collected from patients who anticipate requiring blood transfusion and elect to donate for their own use. Donor-safety screening criteria and testing procedures applicable to blood collection from allogeneic donors do not always apply to these components. Autologous donations must be labeled accordingly, as described below.

Required Testing of Blood Donations

A sample of donor blood is collected at the time of donation and undergoes required testing before labeling and distribution of the associated blood or blood components for routine transfusion. ABO group and Rh type are determined, including testing for the presence of weak D antigen.

All donations intended for transfusion are tested for evidence of relevant transfusion-transmitted infections. Testing is performed by a qualified laboratory using assays that are licensed, approved, or cleared by FDA. Testing on the current donation (or as indicated, below) must indicate that the results are nonreactive for the following:

1. Antibodies to:
 - human immunodeficiency virus, types 1 and 2 (anti-HIV-1/2)
 - hepatitis C virus (anti-HCV)
 - human T-lymphotropic virus, type I and II (anti-HTLV-I/II)

- hepatitis B core antigen (anti-HBc)
 - *Trypanosoma cruzi* either on the current donation or at least one previous donation.
2. Hepatitis B surface antigen (HBsAg).
 3. Licensed nucleic acid test (NAT) for:
 - hepatitis B virus (HBV) deoxyribonucleic acid (DNA)
 - hepatitis C virus (HCV) ribonucleic acid (RNA)
 - human immunodeficiency virus (HIV-1) RNA
 - West Nile virus (WNV) RNA.
 4. Licensed NAT for *Babesia* (RNA and DNA) for blood collected in states where *Babesia* testing is required by FDA, unless pathogen reduction is performed.
 5. Licensed serologic test for *Treponema pallidum* (syphilis).

Warning: The risk of transmitting infectious agents is present. Careful donor selection and available laboratory tests do not eliminate the hazard.

A blood collector may perform additional testing for pathogens; such additional testing may be performed under an FDA-approved investigational new drug (IND) application, using language for component labeling and/or revisions to the *Circular*, as required in the approved IND and provided by the IND sponsor.

Infectious disease testing requirements for autologous donation vary, depending on whether the unit will be drawn in one facility and infused in another facility and whether the units might be made available for allogeneic transfusion. Infectious disease testing is not required for autologous units drawn, stored, and infused at the same facility if the facility does not allow autologous donations to be used for allogeneic transfusion. Autologous units for which testing has not been performed are labeled “DONOR UNTESTED.” Autologous units with reactive test results may be transfused to the donor with appropriate physician authorization. The “BIOHAZARD” label and “FOR AUTOLOGOUS USE ONLY” label will be applied to all autologous units that are tested for evidence of relevant transfusion-transmitted infections as listed above and determined to be reactive. If units labeled “AUTOLOGOUS DONOR” are transfused at a different facility that does not allow autologous donations to be used for allogeneic transfusions, at a minimum the first donation in each 30-day period is tested for evidence of infection as listed above. If testing results are nonreactive, subsequent units not tested within the same 30-day period following initial tests will be labeled as “DONOR TESTED WITHIN THE LAST 30 DAYS.” A “BIOHAZARD” label is required if autologous units have a reactive relevant transfusion-transmitted infection test result within the last 30 days.

In addition, if these units are untested, they must be labeled as “DONOR UNTESTED.” If a facility allows for autologous units to be crossed over for inclusion in the general blood inventory, the donors and units must be subjected to the same donor eligibility and donation suitability requirements and test requirements as allogeneic donors and units.

Tests for unexpected antibodies against red blood cell (red cell) antigens are performed on donor samples for every donation. The results of these tests are negative or have been determined to be clinically insignificant unless otherwise indicated on the label. Other tests may have been performed on donor blood as indicated by information that has been provided by the blood bank or transfusion service on an additional label or tie tag, or in a supplement to this *Circular*.

Bacterial Risk Control Strategies for Platelets

To control the risk of bacterial contamination, platelet components stored at room temperature have been either:

1. tested and found negative for bacterial contamination using FDA-recommended bacterial risk control strategies and FDA-cleared or approved devices, or
2. treated using FDA-approved pathogen reduction technology.

Note: Certain bacterial testing strategies include secondary culture or rapid testing performed prior to transfusion.

Blood and Component Labeling

All **Components Available** identified in this *Circular* are listed using the Information Standard for Blood and Transplant 128 (ISBT 128) product name.

Blood and blood component labels will contain the following information:

1. The proper name, Whole Blood or blood component, including any appropriate qualifications, modifiers, and attributes.
2. The method by which the blood component was prepared, either by whole blood or apheresis collection.
3. The storage temperature range (in degrees Celsius).
4. The preservatives and anticoagulant used in the preparation of the blood or blood components, when appropriate.
5. The standard contents or volume is assumed unless otherwise indicated on the label or in *Circular* supplements.
6. The number of units in pooled blood components.
7. The name, address, registration number, and US license number (if applicable) of the collection and processing location.
8. The expiration date, including the day, month, and year, and, if the dating period for the product is 72 hours or less, including any product prepared in a system that might compromise sterility, the hour of expiration. When the expiration time is not indicated, the product expires at midnight.
9. The donation (unit or pool) identification number.
10. The donor category (paid or volunteer, and autologous, if applicable).
11. ABO group and Rh type, if applicable.
12. Special handling information, as required.
13. Statements regarding proper recipient identification, this *Circular*, infectious disease risk, and prescription requirement.
14. Any sedimenting agent used during cytopheresis, if applicable.

Instructions for Use

The following general instructions pertain to Whole Blood and all the blood components described in this *Circular*:

1. All blood and blood components must be maintained in a controlled environment and stored under appropriate conditions as described in the current version of the AABB *Standards for Blood Banks and Transfusion Services*.
2. The intended recipient and the blood container must be properly identified before the transfusion is started.
3. Aseptic technique must be employed during administration. If the container is entered in a manner that violates the integrity of the system, the component expires 4 hours after entry if maintained at room temperature (20-24 C) or 24 hours after entry if refrigerated (1-6 C).
4. All blood components must be transfused through a sterile, pyrogen-free filter designed to remove clots and aggregates (generally a standard 150- to 260-micron filter).
5. Blood and blood components should be mixed thoroughly before use.
6. Blood and blood components must be inspected immediately before use. If, upon visual inspection, the container is not intact or the appearance is abnormal (presence of excessive hemolysis, a significant color change in the blood bag as compared with the tubing segments, floccular material, cloudy appearance, or other problems), the blood or blood component must not be used for transfusion, and appropriate follow-up with the transfusion service must be performed.
7. No medications or solutions may be added to or infused through the same tubing simultaneously with blood or blood components, with the exception of 0.9% Sodium Chloride, Injection, United States Pharmacopeia (USP), unless: 1) they have been approved for this use by FDA, or 2) there is documentation available to show that the addition is safe and does not adversely affect the blood or blood component.
8. Lactated Ringer's Injection USP or other solutions containing calcium should never be added to or infused through the same tubing with blood or blood components containing citrate.
9. Blood components should be warmed, if clinically indicated, for situations such as exchange or massive transfusions, or for patients with cold-reactive antibodies. Warming must be accomplished using an FDA-cleared warming device.
10. Life-threatening reactions may occur after the infusion of only a small volume of blood or blood components; therefore, unless otherwise indicated by the patient's clinical condition, the rate of infusion should initially be slow.
11. Periodic observation and recording of vital signs should occur before, during, and after the transfusion to identify suspected adverse reactions. If a transfusion reaction occurs, the transfusion must be discontinued

immediately, and appropriate therapy initiated. The infusion should not be restarted unless approved by transfusion service protocol.

12. Specific information and instructions concerning possible adverse reactions shall be provided to the patient or a responsible caregiver when direct medical observation or monitoring of the patient will not be available after transfusion.
13. Transfusion of blood or blood components should start before expiration and finish within 4 hours after entering the container.
14. All adverse events related to transfusion, including possible bacterial contamination of blood or a blood component or suspected disease transmission, must be reported to the transfusion service according to its local protocol.

Refer to the section on **Further Processing** for additional information on:

- Pathogen Reduction Technology
- Leukocyte Reduction
- Irradiation
- Washing and Volume Reduction

Refer to the section on **Additional Testing** for additional information on:

- Identification of CMV-Seronegative Blood
- Identification of Low-Titer Anti-A and/or Anti-B Blood Products

Side Effects and Hazards for Whole Blood and All Blood Components

Transfusion-related adverse events may be voluntarily reported to the National Healthcare Safety Network (NHSN) hemovigilance program (<https://www.cdc.gov/nhsn/biovigilance/index.html>) unless there is a state requirement to report. The NHSN Biovigilance Component Hemovigilance Module Surveillance Protocol (<https://www.cdc.gov/nhsn/pdfs/biovigilance/bv-hv-protocol-current.pdf>) provides case classification criteria for Centers for Disease Control and Prevention-defined transfusion-associated adverse reactions.

Immunologic Complications, Immediate

1. *Hemolytic transfusion reaction*, the immune destruction of red cells, most commonly occurs from exposure of transfused red cells to incompatible recipient plasma. The transfusion of blood components containing plasma which is incompatible with the recipient's red cells rarely results in clinically relevant hemolysis. Further details are discussed in the section on components containing red cells and in the platelets section.
2. *Immune-mediated platelet destruction*, one of the causes of refractoriness to platelet transfusion, is the result of alloantibodies in the recipient to human leukocyte antigen (HLA) or platelet-specific antigens on transfused platelets. This is described in more detail in the section on platelets.
3. *Febrile nonhemolytic reaction* is manifested by a temperature elevation of ≥ 1 C or ≥ 1.8 F or chills/rigors occurring during or within 4 hours after a transfusion and in the absence of any other pyretic stimulus or active warming. This may reflect the action of antibodies against white cells or the action of cytokines either present in the transfused component or generated by the recipient in response to transfused elements. Febrile reactions may occur in less than 1% of transfusions of leukocyte-reduced red cell components and about 5% of leukocyte-reduced apheresis platelet components. Febrile reactions occur more frequently in patients receiving non-leukocyte-reduced components and those previously alloimmunized by transfusion or pregnancy. No routinely available pre- or posttransfusion tests are helpful in predicting or preventing these reactions. Antipyretics usually provide effective symptomatic relief. Patients who experience repeated, severe febrile reactions may benefit from receiving leukocyte-reduced components. If these reactions are caused by cytokines in the component, prestorage leukocyte reduction may be beneficial.
4. *Allergic reactions* frequently occur (ie, with 1-3% of plasma-containing components) as mild or self-limiting urticaria or wheezing that usually responds to antihistamines. More severe manifestations, including respiratory and cardiovascular symptoms, are more consistent with anaphylactoid/anaphylactic reactions and may require more aggressive therapy (see below). No laboratory procedures are available to predict these reactions.

5. *Anaphylactoid/anaphylactic reactions*, characterized by hypotension, tachycardia, nausea, vomiting and/or diarrhea, abdominal pain, severe dyspnea, pulmonary and/or laryngeal edema, and bronchospasm and/or laryngospasm, are rare (<10/100,000 transfused units) but dangerous complications requiring immediate treatment with epinephrine and supportive care. While these reactions have been reported in IgA-deficient patients with anti-IgA antibodies and patients with haptoglobin deficiency, most reactions are idiopathic and not associated with a specific serum protein deficiency, polymorphism, or identifiable cause. In certain circumstances, patients may benefit from the use of washed cellular components to prevent or reduce the severity of allergic reactions not minimized by treatment with medication alone.
6. *Transfusion-related acute lung injury (TRALI)* is characterized by the acute onset of hypoxemia and non-cardiogenic pulmonary edema within 6 hours of a blood or blood component transfusion not attributable to other causes of acute lung injury or circulatory overload. Various stimuli in blood components, most commonly white cell antibodies from donors sensitized during pregnancy or prior transfusion or transplantation, or proinflammatory molecules that accumulate in stored blood components, may cause TRALI. These mechanisms may not be mutually exclusive and may act synergistically with underlying patient factors to lead to a final common pathway of acute lung injury. These stimuli may trigger an inflammatory response, granulocyte activation and degranulation, and injury to the alveolar capillary membrane and the development of permeability pulmonary edema. Although most TRALI cases are associated with donor antileukocyte antibodies, rare cases have implicated recipient antileukocyte antibodies that reacted with donor leukocytes. Widespread leukoreduction of blood components has likely mitigated this latter risk. Laboratory testing of blood donors for antileukocyte antibodies or blood components for biological mediators does not alter management of this reaction, which is diagnosed on clinical and radiographic findings. Treatment of TRALI involves aggressive respiratory support, and often mechanical ventilation. The preferential use of plasma collected from male donors or female donors who have tested negative for the presence of HLA Class I and/or II antibodies has been associated with a significant reduction in the number of reported TRALI cases and associated fatalities. Transfusion services should immediately report suspected TRALI to the blood collection facility to facilitate the retrieval of other components associated with the involved donation(s) or prior donations.

Immunologic Complications, Delayed

1. *Delayed hemolytic reaction* is described in detail in the sections on components containing red cells.
2. *Alloimmunization* to antigens of red cells, white cells, platelets, or plasma proteins may occur unpredictably after transfusion. Blood components may contain certain immunizing substances other than those indicated on the label. For example, platelet components may also contain red cells and white cells. Primary immunization does not become apparent until days or weeks after the immunizing event and does not usually cause symptoms or physiologic changes. If components that express the relevant antigen are subsequently transfused, there may be accelerated removal of cellular elements from the circulation and/or systemic symptoms. Clinically significant antibodies to red cell antigens will ordinarily be detected by pretransfusion testing. Alloimmunization to antigens of white cells, platelets, or plasma proteins can be detected only by specialized testing.
3. *Posttransfusion purpura* is a rare syndrome characterized by the development of dramatic, sudden, and self-limited thrombocytopenia, typically 7 to 10 days after a blood transfusion, in a patient with a history of sensitization by either pregnancy or transfusion. Although the immune specificity may be to a platelet-specific antigen the patient lacks, both autologous and allogeneic platelets are destroyed. High-dose Immune Globulin, Intravenous (IVIG) may correct the thrombocytopenia.
4. *Transfusion-associated graft-vs-host disease (TA-GVHD)* is rare but has a fatality rate of nearly 100% due to overwhelming infection in the setting of pancytopenia. This condition occurs when viable T lymphocytes in the transfused component engraft in the recipient and react against recipient tissue antigens. TA-GVHD can occur if the host does not recognize and reject the foreign transfused cells, and it can follow transfusion of any component that contains even very small numbers of viable T lymphocytes. Immunologically normal recipients who are heterozygous for a tissue antigen haplotype for which the donor is homozygous are at risk. Recipients with severe cellular immunodeficiency (except for HIV infection) are also at greatest risk (eg, fetuses receiving intrauterine transfusions, at-risk neonates, recipients of hematopoietic progenitor cell transplants, and selected patients with severe immunodeficiency conditions). Patients with oncologic and rheumatologic diseases receiving purine analogues (eg, fludarabine, cladribine) or certain other biological immunomodulators (eg, alemtuzumab, antithymocyte globulin) may be at risk for TA-GVHD, depending

on clinical factors and the source of the biological agent. TA-GVHD remains a risk with leukocyte-reduced components because they contain sufficient residual T lymphocytes. Irradiation of the component renders T lymphocytes incapable of proliferation. Pathogen reduction technology may also be used as an alternative to irradiation to prevent TA-GVHD if the pathogen reduction technology has been shown to inactivate residual lymphocytes.

Nonimmunologic Complications

Because Whole Blood and blood components are made from human blood, they may carry a risk of transmitting infectious agents [eg, viruses, bacteria, parasites, the variant Creutzfeldt-Jakob disease (vCJD) agent, and, theoretically, the Creutzfeldt-Jakob disease agent (CJD)]. Also, septic and toxic reactions can result from transfusion of bacterially contaminated blood and blood components. Careful donor selection, available laboratory tests, and pathogen reduction technology do not totally eliminate these hazards. Such complications are infrequent but may be life-threatening. Infectious disease transmission may occur despite careful selection of donors and testing of blood. Donor selection criteria are designed to screen out potential donors with increased risk of infection with HIV, HTLV, hepatitis, and syphilis, as well as other agents (see section on Required Testing of Blood Donations). For *other infectious agents* (eg, *the vCJD agent*), there are **no** licensed tests available for donor testing; however, other screening measures for possible exposure or history of vCJD, CJD, or use of pathogen reduction technology may mitigate the risk of transfusion-transmitted infections. Transfusion services should immediately report infections that may be related to the blood donor or to the manufacturing of blood components to the collection facility.

1. *Cytomegalovirus (CMV)* may be present in white-cell-containing components from donors previously infected with this virus, which can persist for a lifetime despite the presence of serum antibodies. Up to 70% of donors may be CMV seropositive. Transmission of CMV by transfusion may be of concern in low-birthweight (≤ 1200 g) premature infants born to CMV-seronegative mothers and in intrauterine transfusions and/or certain other categories of immunocompromised individuals such as hematopoietic progenitor cell or solid-organ transplant patients if they are CMV seronegative. For at-risk recipients, the risk of CMV transmission by cellular components can be reduced by transfusing CMV-seronegative or leukocyte-reduced components, or pathogen-reduced components when applicable.
2. *Bacterial sepsis* occurs rarely but can cause acute, severe, sometimes life-threatening effects. Onset of high fever (≥ 2 C or ≥ 3.5 F increase in temperature), severe chills, hypotension, or circulatory collapse during or shortly after transfusion should suggest the possibility of bacterial contamination and/or endotoxin reaction in the transfused products. Although platelet components stored at room temperature have been implicated most frequently, previously frozen components thawed by immersion in a water bath and red cell components stored for several weeks at 1 to 6 C have also been implicated. Platelet components are controlled for bacterial contamination; however, this does not completely eliminate the risk.

Both gram-positive and gram-negative organisms have been identified as causing septic reactions. Organisms capable of multiplying at low temperatures (eg, *Yersinia enterocolitica*) and those using citrate as a nutrient have been associated with components containing red cells. A variety of pathogens, as well as skin contaminants, have been found in platelet components. Multiplication of gram-negative bacteria in blood components has also caused endotoxemia in recipients.

Prompt recognition of a possible septic reaction is essential, with immediate discontinuation of the transfusion and aggressive therapy with broad-spectrum antimicrobials and vasopressor agents, if necessary. In addition to prompt sampling of the patient's blood for cultures, investigation should include examination of material from the blood container by Gram stain, and cultures of specimens from the container and the administration set. It is important to report all febrile transfusion reactions to the transfusion service for appropriate investigation. If posttransfusion sepsis is suspected, the transfusion service should immediately report the reaction to the blood collection facility to facilitate retrieval of other potentially contaminated components associated with the collection.

3. *Transfusion-associated circulatory overload (TACO)* is a frequent complication of transfusion, leading to cardiogenic (hydrostatic) pulmonary edema and can occur after transfusion of excessive volumes of blood components or transfusing at excessively rapid rates. Signs and symptoms include new or worsening respiratory distress and radiographic and/or clinical evidence of volume overload. Individuals with underlying cardiopulmonary or renal disease, the very young and the elderly, and patients with chronic severe anemia in whom low red cell mass is associated with high plasma volume are at particular risk. Small transfusion volumes can precipitate symptoms in at-risk patients who already have a positive fluid balance.

Pulmonary edema should be promptly and aggressively treated, and infusion of colloid preparations, including plasma components and the supernatant fluid in cellular components, reduced to a minimum.

4. *Hypothermia* carries a risk of cardiac arrhythmia or cardiac arrest and exacerbation of coagulopathy. Rapid infusion of large volumes of cold blood or blood components can depress body temperature, and the danger is compounded in patients experiencing shock or surgical or anesthetic manipulations that disrupt temperature regulation. A blood warming device should be considered if rapid infusion of blood or blood components is needed. Warming must be accomplished using an FDA-cleared blood warming device so as not to cause hemolysis.
5. *Metabolic complications* may accompany large-volume transfusions, especially in neonates and patients with liver or kidney disease.
 - a. Citrate “toxicity” reflects a depression of ionized calcium caused by the presence in the circulation of large quantities of citrate anticoagulant. Because citrate is promptly metabolized by the liver, this complication is rare. Patients with severe liver disease or those with circulatory collapse that prevents adequate hepatic blood flow may have physiologically significant hypocalcemia after rapid, large-volume transfusion. Citrated blood or blood components administered rapidly through central intravenous access may reach the heart so rapidly that ventricular arrhythmias occur. Standard measurement of serum calcium does not distinguish ionized from complexed calcium. Ionized calcium testing or electrocardiogram monitoring is more helpful in detecting physiologically significant alteration in calcium levels.
 - b. Other metabolic derangements can accompany rapid or large-volume transfusions, especially in patients with preexisting circulatory or metabolic problems. These include acidosis or alkalosis (deriving from changing concentrations of citric acid and its subsequent conversion to pyruvate and bicarbonate) and hyper- or hypokalemia.

Fatal Transfusion Reactions

Reporting requirements can be found on the FDA webpage, *Transfusion/Donation Fatalities*:

“Section 606.170(b) of Title 21, Code of Federal Regulations [21 CFR 606.170(b)], requires that facilities notify the Food and Drug Administration (FDA), Center for Biologics Evaluation and Research (CBER), Office of Compliance and Biologics Quality (OCBQ), as soon as possible after confirming a complication of blood collection or transfusion to be fatal. The collecting facility is to report donor fatalities, and the compatibility testing facility is to report recipient fatalities. The regulation also requires the reporting facility to submit a report of the investigation within 7 days after the fatality.”

FDA’s August 2021 *Guidance, Notifying FDA of Fatalities Related to Blood Collection or Transfusion; Guidance for Industry*, provides recommendations and additional information, including this clarification:

“We recommend that you submit the initial notification by email, if possible, and if you do so, you will receive an email confirmation receipt from us. If email is not feasible, please notify us by telephone or facsimile. We cannot access notification outside of customary working hours unless you use email or telephone.”

When reporting a fatality during or outside of regular business hours, the reporting facility may submit initial notification by leaving a voice message or sending an email or facsimile to the Division of Inspections and Surveillance.

- Email: fatalities2@fda.hhs.gov
- Telephone/voice-mail number: 240-402-9160
- Fax number: 301-837-6256, Attn: CBER Fatality Program Manager
- Express mail address: See below.

FDA will contact you as soon as possible to obtain more detailed information. This does not replace the 7-day written report regarding the fatality and all related information as described in 21 CFR 606.170(b).

The 7-day follow-up report may be submitted by email, facsimile, or express mail.

Express mail address for 7-day follow-up reports:

US Food and Drug Administration
 Center for Biologics Evaluation and Research Document Control Center
 10903 New Hampshire Avenue
 WO71, G112
 Silver Spring, MD 20993-0002

Refer to FDA's website for information (<https://www.fda.gov/vaccines-blood-biologics/report-problem-center-biologics-evaluation-research/transfusion-donation-fatalities>) and the August 2021 Guidance for Industry, *Notifying FDA of Fatalities Related to Blood Collection or Transfusion*.

Whole Blood

Overview

Whole Blood is transfused to increase oxygen-carrying capacity in patients whose physiologic compensatory mechanisms are inadequate to maintain normal tissue oxygenation. Whole Blood may be transfused in an emergency situation or other clinical setting that necessitates delivery of multiple blood components simultaneously. When preservation of platelet function is required, Whole Blood intended for transfusion should be collected from a donor who has not recently ingested a drug that adversely affects platelet function. (Refer to the current version of the AABB Donor History Questionnaire and the Medication Deferral List.)

Description

A single whole blood donation typically contains either 450 milliliters (mL) ($\pm 10\%$) or 500 mL ($\pm 10\%$) of blood collected from allogeneic blood donors with a minimum hematocrit of 36% or 38% (females) or 39% (males), drawn in a sterile container that includes an anticoagulant solution licensed for this component. Whole Blood is prepared in an aseptic manner in a ratio of 14 mL of anticoagulant-preservative solution per 100 mL of whole blood targeted for collection.

Whole Blood contains approximately 5.5×10^{10} platelets. The volume of plasma in Whole Blood is about 170 mL or greater and contains nonlabile clotting factors.

Whole Blood must be stored at 1 to 6 C for an interval ("shelf life") determined by the properties of the anticoagulant-preservative solution (see Table 1).

Refer to the section on **Further Processing** for additional information on:

- Leukocyte Reduction

Refer to the section on **Additional Testing** for additional information on:

- Identification of CMV-Seronegative Blood
- Identification of Low-Titer Anti-A and/or Anti-B Blood Products

Actions

Whole Blood increases the recipient's oxygen-carrying capacity by increasing the mass of circulating red cells. In addition to red cells, Whole Blood provides plasma and platelets, which provide volume expansion and may contribute to hemostasis.

Indications

Whole Blood may be indicated in life-threatening hemorrhage where oxygen-carrying capacity, nonlabile coagulation factors, platelets, and volume expansion are needed.

Contraindications

Whole Blood should not be used solely for volume expansion or to increase oncotic pressure of circulating blood.

Dosage and Administration

Whole Blood contains enough hemoglobin to increase the hemoglobin concentration in an average-sized adult by approximately 1 gram/deciliter (g/dL) (increase hematocrit by 3%).

Whole Blood must be ABO group-specific with the recipient. Serologic compatibility between recipient and donor should be established when any red-cell-containing component is transfused. This may be accomplished by performing ABO/Rh typing, antibody screening, and crossmatching by serologic technique or use of a computer crossmatch. In life-threatening situations, group O Whole Blood may be administered to non-O patients, provided facilities have policies and procedures to define titer cutoffs for anti-A and anti-B titers.

The transfusing facility must have policies and procedures in place addressing specific indications for use, product specifications, administration instructions, and a defined maximum number of units to be transfused to each patient.

Table 1. Contents of Anticoagulant-Preservative Solutions*

Anticoagulant-Preservative (g/L)	Trisodium Citrate	Citric Acid	Monobasic Sodium Phosphate	Dextrose	Adenine	Shelf Life(Days)
Anticoagulant citrate-dextrose A (ACD-A) [†]	22.0	8.0	0	24.5	0	21
Citrate-phosphate-dextrose (CPD)	26.3	3.27	2.22	25.5	0	21
Citrate-phosphate-dextrose-dextrose (CP2D)	26.3	3.27	2.22	51.1	0	21
Citrate-phosphate-dextrose-adenine (CPDA-1)	26.3	3.27	2.22	31.9	0.275	35

*63 mL/450 mL collection, 70 mL/500 mL collection.

[†]ACD is used for apheresis components.

Table 2. Contents of Red Blood Cells Additive Solutions*

Additive Solution (mg/100 mL)	Dextrose Monohydrate	Adenine	Monobasic Sodium Phosphate	Dibasic Sodium Phosphate	Mannitol	Sodium Bicarbonate	Sodium Chloride	Sodium Citrate	Citric Acid	Shelf Life (Days)
AS-1 (Adsol)	2200	27	0	0	750	0	900	0	0	42
AS-3 (Nutricel)	1100	30	276	0	0	0	410	588	42	42
AS-5 (Optisol)	900	30	0	0	525	0	877	0	0	42
AS-7 (SOLX)	1585	27	0	170	1000	218	0	0	0	42

*100 mL AS/450-mL collection, 110 mL AS/500-mL collection.

The initial portion of each unit transfused should be infused cautiously and with sufficient observation to detect onset of acute reactions. Thereafter, the rate of infusion can be more rapid, as tolerated by the patient's circulatory system. It is undesirable for components that contain red cells to remain at room temperature longer than 4 hours.

Side Effects and Hazards

Hazards that pertain to all transfusion components are described in the earlier section titled Side Effects and Hazards for Whole Blood and All Blood Components. Listed below is additional information on hazards that apply specifically to components that contain red cells.

1. **Hemolytic transfusion reaction** is the immunologic destruction of transfused red cells, nearly always the result of incompatibility of antigen on the transfused cells with antibody in the recipient's circulation (see item 4 below for discussion of nonimmunologic hemolysis). The most common cause of severe, acute hemolytic reactions is transfusion of ABO-incompatible blood, resulting from identification errors occurring at some point(s) in the transfusion process. Serologic incompatibility undetected during pretransfusion testing is a much less common cause of acute hemolysis. If a hemolytic transfusion reaction is suspected, the transfusion must be stopped, and the transfusion service laboratory notified immediately. Information identifying the patient and the transfusion component, and associated forms and labels must be reviewed promptly to detect possible errors. A postreaction blood sample, preferably drawn from a site other than the transfusion access, must be sent to the laboratory along with the implicated unit of blood and administration set.

Acute hemolytic reactions characteristically begin with an increase in temperature and pulse rate; symptoms may include chills, dyspnea, chest or back pain, abnormal bleeding, or shock. Instability of blood pressure is frequent, the direction and magnitude of change depending upon the phase of the reaction and the magnitude of compensatory mechanisms. In anesthetized patients, hemoglobinuria, hypotension, and evidence of disseminated intravascular coagulopathy (DIC) may be the first signs of incompatibility. Laboratory findings can include hemoglobinemia and/or hemoglobinuria, followed by elevation of serum indirect bilirubin. The direct antiglobulin test (DAT) result is usually positive, with rare exceptions (ie, complete hemolysis of incompatible red cells). Treatment includes measures to maintain or correct arterial blood pressure; correct coagulopathy, if present; and promote or maintain renal function. Lack of symptoms does not exclude an acute hemolytic reaction.

Delayed hemolytic reactions occur in previously red-cell-alloimmunized patients in whom antigens on transfused red cells provoke anamnestic production of antibody. The anamnestic response reaches a significant circulating level while the transfused cells are still present in the circulation; the usual time frame is 2 to 14 days after transfusion. Signs may include unexplained fever, development of a positive DAT result, and unexplained decrease in hemoglobin/hematocrit. Hemoglobinemia and hemoglobinuria are uncommon, but elevation of lactate dehydrogenase or bilirubin may be noted. Most delayed hemolytic reactions have a benign course and require no treatment.

Hemolytic transfusion reactions in patients with sickle cell anemia may be particularly severe, with destruction of autologous as well as transfused red cells, resulting in a lower hemoglobin level after transfusion. This is suggestive of hyperhemolysis syndrome. In such patients, serologic investigations may not reveal the specificity of the causative antibody. Immediate treatment may include steroid use, IVIG, and avoiding transfusions, if possible. Consultation with a transfusion medicine specialist is required in these cases. Prospective matching for Rh and Kell antigens may decrease risk.

2. Antigens on transfused red cells may cause red cell **alloimmunization** of the recipient. Clinically significant antibodies to red cell antigens will usually be detected in pretransfusion antibody screening tests. For most patients, red cell antigen matching beyond ABO and Rh is unnecessary.
3. **TACO** can accompany transfusion of any component at a rate more rapid than the recipient's cardiac output can accommodate. Whole Blood creates more of a risk than Red Blood Cell components (RBCs) because the transfused plasma adds volume without increasing oxygen-carrying capacity. Patients with chronic anemia have increased plasma volumes and are at increased risk for circulatory overload.
4. **Nonimmunologic hemolysis** occurs rarely but can result from: 1) introduction of hypotonic fluids into the circulation; 2) effects of drugs coadministered with transfusion; 3) effects of bacterial toxins; 4) thermal injury by freezing or overheating; 5) metabolic damage to cells, as from hemoglobinopathies or enzyme deficiencies; or 6) mechanical injury or osmotic stresses. Examples of situations capable of causing nonimmune red cell hemolysis include exposure to excessive heat when using warming devices not cleared or approved

by FDA, mixture of blood with hypotonic solutions, or transfusion under high pressure through small-gauge or defective needles.

Components Available

WHOLE BLOOD is prepared from 400-550 mL of blood collected into the appropriate volume of anticoagulant solution.

WHOLE BLOOD LEUKOCYTES REDUCED is prepared from Whole Blood by a method resulting in a final product containing $<5.0 \times 10^6$ leukocytes and $\geq 85\%$ of the original Whole Blood content. Leukocyte-reduced Whole Blood may be prepared using a platelet-sparing leukocyte reduction filter.

Red Blood Cell Components

Overview

RBCs are transfused to increase oxygen-carrying capacity in patients whose physiologic compensatory mechanisms are inadequate to maintain normal tissue oxygenation. Red cells contain hemoglobin and serve as the primary agent for transport of oxygen to tissues. The primary red-cell-containing transfusion component is RBCs. This component is prepared by centrifugation or sedimentation of Whole Blood to remove much of the plasma. RBC components can also be prepared by apheresis methods.

Description

Depending upon the collection system used, a single whole blood donation typically contains either 450 mL ($\pm 10\%$) or 500 mL ($\pm 10\%$) of blood collected from allogeneic blood donors with a minimum hematocrit of 36% to 38% (females) or 39% (males), withdrawn in a sterile container that includes an anticoagulant solution licensed for this component. In the case of autologous adult blood donors, a hematocrit minimum as low as 33% is acceptable. Occasionally, units of other volumes are collected, and those volumes are stated on the label.

Red-cell-containing components can be stored at 1 to 6 C for an interval ("shelf life") determined by the properties of the anticoagulant-preservative solution (see Table 1). Whole Blood units are prepared in an aseptic manner in a ratio of 14 mL of anticoagulant-preservative solution per 100 mL of Whole Blood targeted for collection. Apheresis components are collected into anticoagulants as recommended by the manufacturer.

After plasma is removed, the resulting component is RBCs, which has a hematocrit between 65% and 80% and a usual volume between 225 mL and 350 mL. RBC additive solutions (AS) may be mixed with the red cells remaining after removal of nearly all of the plasma to extend the shelf life (see Table 2). The typical hematocrit of AS RBCs is 55% to 65%, and the volume is approximately 300 to 400 mL. AS RBCs have a shelf life of 42 days. Descriptions of specific components containing red cells are given at the end of this section.

Refer to the section on **Further Processing** for additional information on:

- Leukocyte Reduction
- Irradiation
- Washing and Volume Reduction

Refer to the section on **Additional Testing** for additional information on:

- Identification of CMV-Seronegative Blood
- Identification of Low-Titer Anti-A and/or Anti-B Blood Products

Actions

RBC components increase the recipient's oxygen-carrying capacity by increasing the mass of circulating red cells. Processing and/or storage deplete the component of virtually all potential therapeutic benefit attributable to the functions of white cells and platelets; however, cellular elements remain in these blood components and may cause adverse immunologic or physiologic consequences. Residual plasma in the component provides the recipient with volume expansion and nonlabile plasma proteins to the extent that residual plasma is present in the preparation. Depending on the method of production, RBCs may contain approximately 20 to 100 mL of residual plasma. RBCs prepared with AS are the most used red cell product and have limited residual plasma.

Indications

Red-cell-containing components are indicated for treatment of symptomatic or critical deficit of oxygen-carrying capacity. They are also indicated for red cell exchange transfusion.

Contraindications

Red-cell-containing components should not be used to treat anemias that can be corrected with specific hematinic medications such as iron, vitamin B12, folic acid, or erythropoietin.

RBCs should not be used solely for volume expansion or to increase oncotic pressure of circulating blood.

Dosage and Administration

Each unit of RBCs contains enough hemoglobin to increase the hemoglobin concentration in an average-sized adult by approximately 1 g/dL (increase hematocrit by 3%). Smaller aliquots can be made available for use with neonatal or pediatric patients, or adults with special transfusion needs.

The ABO group of all red-cell-containing components must be compatible with ABO antibodies in the recipient's plasma.

Serologic compatibility between recipient and donor must be established before any red-cell-containing component is transfused. This may be accomplished by performing ABO/Rh typing, antibody screening, and crossmatching by serologic technique or use of a computer crossmatch. Incases when delay in transfusion will be life-threatening, uncrossmatched group O RBCs or ABO group-specific RBCs may be transfused before completion of pretransfusion compatibility testing.

The initial portion of each unit transfused should be infused cautiously and with sufficient observation to detect onset of acute reactions. Thereafter, the rate of infusion can be more rapid, as tolerated by the patient's circulatory system. It is undesirable for components that contain redcells to remain at room temperature longer than 4 hours. If the anticipated infusion rate must be so slow that the entire unit cannot be infused within 4 hours, it is appropriate to order smaller aliquots for transfusion.

See Table 3 for pediatric dosage information.

Side Effects and Hazards

Hazards that pertain to all transfusion components are described in the earlier section titled Side Effects and Hazards for Whole Blood and All Blood Components. Listed below are hazards that apply specifically to components that contain red cells.

1. **Hemolytic transfusion reaction** is the immunologic destruction of transfused red cells, nearly always the result of incompatibility of antigen on the transfused cells with antibody in the recipient's circulation (see item 5 below for discussion of nonimmunologic hemolysis). The most common cause of severe, acute hemolytic reactions is transfusion of ABO-incompatible blood, resulting from identification errors occurring at some point(s) in the transfusion process. Serologic incompatibility undetected during pretransfusion testing is a much less common cause of acute hemolysis. If a hemolytic transfusion reaction is suspected, the transfusion must be stopped, and the transfusion service laboratory notified immediately. Information identifying the patient, the transfusion component, and associated forms and labels must be reviewed promptly to detect possible errors. A postreaction blood sample, preferably drawn from a site other than the transfusion access, must be sent to the laboratory along with the implicated unit of blood and administration set.

Acute hemolytic reactions characteristically begin with an increase in temperature and pulse rate; symptoms may include chills, dyspnea, chest or back pain, abnormal bleeding, or shock. Instability of blood pressure is frequent, the direction and magnitude of change depending upon the phase of the reaction and the magnitude of compensatory mechanisms. In anesthetized patients, hemoglobinuria, hypotension, and evidence of DIC may be the first signs of incompatibility. Laboratory findings can include hemoglobinemia and/or hemoglobinuria, followed by elevation of serum indirect bilirubin. The DAT result is usually positive, with rare exceptions (ie, complete hemolysis of incompatible red cells). Treatment includes measures to maintain or correct arterial blood pressure; correct coagulopathy, if present; and promote or maintain renal function. Lack of symptoms does not exclude an acute hemolytic reaction.

Delayed hemolytic reactions occur in previously red-cell-alloimmunized patients in whom antigens on transfused red cells provoke anamnestic production of antibody. The anamnestic response reaches a significant circulating level while the transfused cells are still present in the circulation; the usual time frame is 2 to 14 days after transfusion. Signs may include unexplained fever, development of a positive DAT result, and unexplained decrease in hemoglobin/hematocrit. Hemoglobinemia and hemoglobinuria are uncommon, but elevation of lactate dehydrogenase or bilirubin may be noted. Most delayed hemolytic reactions have a benign course and require no treatment.

Hemolytic transfusion reactions in patients with sickle cell anemia may be particularly severe, with destruction of autologous as well as transfused red cells, resulting in a lower hemoglobin level after transfusion. This is suggestive of hyperhemolysis syndrome. In such patients, serologic investigations may not reveal the specific-

Table 3. Suggested Pediatric (patients <50 kg) Dosing (adapted from Mo Y, Roseff SD, Wong ECC, eds. Pediatric hemotherapy data card. 5th ed. Bethesda, MD: AABB, 2020)

Component	Attributes	Dosage	Expected Increment
Red Blood Cells (RBCs)	CPD, CPDA-1 (65-80% Hct)	5-15 mL/kg	3 g/dL rise in Hb
	AS-1, AS-3, AS-5, AS-7 (55-65% Hct)	10-15 mL/kg	2 g/dL rise in Hb
Washed RBCs*	70-80% Hct, suspended in normal saline	10-15 mL/kg	3 g/dL rise in Hb
Plasma components [†]	Near-normal levels of coagulation factors, citrate anticoagulant	10-15 mL/kg	15-20% rise in factor level (assume ideal recovery)
Platelets	Whole-blood-derived platelets: $\geq 5.5 \times 10^{10}$ platelets suspended in 25-50 mL of plasma	5-10 mL/kg OR 1 unit/10 kg (patients >10 kg)	50,000-100,000/ μ L rise in platelet count (assume ideal recovery)
	Apheresis platelets: $\geq 3.0 \times 10^{11}$ platelets in 250-300 mL plasma or platelet additive solution equivalent to approx. 6 units of whole-blood-derived platelets	Same as above	Same as above (assume ideal recovery)
Cryoprecipitated AHF	≥ 150 mg fibrinogen/single donor, ≥ 80 units Factor VIII/unit, von Willebrand Factor (vWF), Factor XIII	1-2 units/10 kg (volume of a unit will vary, maximum to 15 mL) or 2-3 mL/kg	60-100 mg/dL rise in fibrinogen
Granulocytes [‡]	Apheresis or pooled from whole blood buffy coats	10-15 mL/kg (1×10^9 to 2×10^9 polymorphonuclear cells/kg) for neonates. For older children, minimum of 1×10^{10} granulocytes.	None. Administered daily until an adequate neutrophil count is maintained and/or the patient shows clinical improvement.

*Wong ECC, Roseff SD, Bandarenko N, eds. Pediatric hemotherapy data card. 4th ed. Bethesda, MD: AABB, 2015.

[†]See Table 7 for specific components.

[‡]Roseff SD, Luban NL, Manno CS. Guidelines for assessing appropriateness of pediatric transfusion. *Transfusion* 2002;42:1398-413; Wong ECC, Roseff SD, Bandarenko N, eds. Pediatric transfusion: A handbook. 5th ed. Bethesda, MD: AABB, 2020; Price TH, Boeckh M, Harrison RW, et al. Efficacy of transfusion with granulocytes from G-CSF/dexamethasone-treated donors in neutropenic patients with infection. *Blood* 2015;126:2153-61; Goel R, Punzalan RC, Wong ECC. Neonatal and pediatric transfusion practice. In: Cohn CS, Delaney M, Johnson ST, et al, eds. Technical manual. 21st ed. Bethesda, MD: AABB, 2023:620.

AHF = antihemophilic factor; AS = additive solution; CPD = citrate-phosphate-dextrose; CPDA = citrate-phosphate-dextrose-adenine; Hb = hemoglobin; Hct = hematocrit.

ity of the causative antibody. Immediate treatment may include steroid use, IVIG, and avoiding transfusions, if possible. Consultation with a transfusion medicine specialist is required in these cases. Prospective matching for Rh and Kell antigens may decrease risk.

2. Antigens on transfused red cells may cause red cell **alloimmunization** of the recipient. Clinically significant antibodies to red cell antigens will usually be detected in pretransfusion antibody screening tests. For most patients, red cell antigen matching beyond ABO and Rh is unnecessary.
3. **TACO** can accompany transfusion of any component at a rate more rapid than the recipient's cardiac output can accommodate. Patients with chronic anemia have increased plasma volumes and are at increased risk for circulatory overload.
4. **Iron overload** is a complication of chronic RBC transfusion therapy. Each transfusion contributes approximately 250 milligrams (mg) of iron, and significant accumulation can occur after 10 to 20 RBC transfusions. Patients requiring multiple transfusions due to decreased red cell production or increased RBC destruction are at far greater risk than patients transfused for hemorrhagic indications, because blood loss is an effective means of iron excretion. Patients with predictably chronic transfusion requirements should be considered for treatment with iron-chelating agents, a program of exchange transfusion therapy, or therapeutic phlebotomy, if applicable.
5. **Nonimmunologic hemolysis** occurs rarely but can result from: 1) introduction of hypotonic fluids into the circulation; 2) effects of drugs coadministered with transfusion; 3) effects of bacterial toxins; 4) thermal injury by freezing or overheating; 5) metabolic damage to cells, as from hemoglobinopathies or enzyme deficiencies; or 6) mechanical injury or osmotic stresses. Examples of situations capable of causing nonimmune red cell hemolysis include exposure to excessive heat when using warming devices not cleared or approved by FDA, mixture of blood with hypotonic solutions, or transfusion under high pressure through small-gauge or defective needles.

Components Available

RED BLOOD CELLS are prepared from blood collected into any of the anticoagulant-preservative solutions approved by the FDA and separated from the plasma by centrifugation or sedimentation. Separation may be done at any time during the allowable shelf life. RBCs may contain from 160 to 275 mL of red cells [50-80 grams (g) of hemoglobin] suspended in varying quantities of residual plasma.

RED BLOOD CELLS ADENINE SALINE ADDED are prepared by centrifuging Whole Blood to remove as much plasma as possible and replacing the plasma with usually 100 to 110 mL of an AS that contains some combination (see Table 2) of dextrose, adenine, sodium chloride, sodium bicarbonate, monobasic or dibasic sodium phosphate, or mannitol; the hematocrit is usually between 55% and 65%. RBCs in an AS have lower viscosity than RBCs, and flow through administration systems in a manner more comparable to that of Whole Blood. RBCs stored with an AS have an extended shelf life.

RED BLOOD CELLS LEUKOCYTES REDUCED and RED BLOOD CELLS ADENINE SALINE ADDED LEUKOCYTES REDUCED are prepared from a unit of Whole Blood (collected in anticoagulant-preservative solution as noted above) containing ≥ 1 to 10×10^9 white cells. In general, leukocyte reduction is achieved by filtration: 1) soon after collection (prestorage) or 2) after varying periods of storage in the laboratory. Leukocyte reduction will decrease the cellular content and volume of blood according to characteristics of the filter system used. RBCs Leukocytes Reduced must have a residual content of leukocytes $< 5.0 \times 10^6$. Leukocyte reduction filters variably remove other cellular elements in addition to white cells. The leukocyte-reduced component contains $\geq 85\%$ of the original red cell content.

RED BLOOD CELLS, ADENINE SALINE ADDED, LEUKOCYTES REDUCED, O₂/CO₂ REDUCED are prepared following collection and processing of red blood cells leukocytes reduced in a processing system that limits O₂ and CO₂ levels in the storage environment. The component may be stored under reduced oxygen conditions for up to 42 days at 1 to 6 C.

APHERESIS RED BLOOD CELLS are red cells collected by apheresis. This component must be collected in an approved anticoagulant. The red cell volume collected and the anticoagulant used are noted on the label. Aside from the automated collection method used, the component is comparable to whole-blood-derived RBCs in all aspects. The dose can be calculated, as for RBCs, from the red cell content of the product. Apheresis RBCs contain approximately 60 g of hemoglobin per unit.

APHERESIS RED BLOOD CELLS LEUKOCYTES REDUCED and APHERESIS RED BLOOD CELLS ADENINE SALINE ADDED LEUKOCYTES REDUCED are collected by apheresis methods.

Leukocyte reduction is achieved by filtration during the manufacturing process resulting in a final product containing $<5.0 \times 10^6$ leukocytes and $\geq 85\%$ of the target red cell content.

RED BLOOD CELLS, LOW VOLUME are prepared when 300 to 404 mL of Whole Blood is collected into an anticoagulant volume calculated for 450 mL \pm 45 mL or when 333 to 449 mL of Whole Blood is collected into an anticoagulant volume calculated for 500 mL \pm 50 mL. These products reflect a collection with an altered ratio of anticoagulant to red cells and may not be an indication of a lower dose of hemoglobin. Plasma and platelet components should not be prepared from low-volume collections.

FROZEN RED BLOOD CELLS and FROZEN REJUVENATED RED BLOOD CELLS are prepared by adding glycerol to red cells as a cryoprotective agent before freezing at -65 C or colder. The glycerol must be removed from the thawed component before it is infused. Frozen RBCs can be stored for up to 10 years. Some rare units may be stored frozen beyond 10 years, provided there is exceptional medical need for the units. Frozen storage is especially suitable for red cells with unusual antigenic phenotypes.

DEGLYCEROLIZED RED BLOOD CELLS is the form in which cryopreserved red cells (Frozen RBCs) are made available for infusion. Glycerol is added to red cells as a cryoprotective agent before freezing and must be removed from the thawed component before it is infused.

Deglycerolized RBCs contain 80% or more of the red cells present in the original unit of blood and have approximately the same expected posttransfusion survival as RBCs. Glycerol is removed by washing the cells with progressively lower concentrations of Sodium Chloride, Injection USP; the final suspension is in 0.9% Sodium Chloride, Injection USP, with or without small amounts of dextrose. Small amounts of residual-free hemoglobin may cause the supernatant fluid to be pink-tinged.

Deglycerolized RBCs provide the same physiologic benefits as RBCs, but their use is usually restricted to situations in which standard transfusion components are inappropriate or unavailable. Deglycerolized RBCs may be useful for transfusions to patients with previous severe allergic transfusion reactions because the process efficiently removes plasma constituents.

In addition to the side effects and hazards of RBC transfusion, Deglycerolized RBCs carry a risk of intravascular hemolysis if deglycerolization has been inadequate.

Deglycerolized RBCs must be transfused within 24 hours after thawing if prepared in an open system. If prepared in a closed system, they can be stored at 1 to 6 C and infused within a 2-week interval after thawing and as directed by the manufacturer's instructions for use.

REJUVENATED RED BLOOD CELLS may be prepared from red cells stored at 1 to 6 C and prepared with CPD and CPDA-1 up to 3 days after expiration. RBCs stored in CPD/AS-1 or CP2D/AS-3 may be rejuvenated up to, but not exceeding 42 days of uninterrupted storage at 1 to 6 C. Addition of an FDA-approved solution containing inosine, phosphate, and adenine restores 2,3-diphosphoglycerate and adenosine triphosphate to levels approximating those of freshly drawn cells. These products must be washed before infusion to remove the inosine, which may be toxic. Rejuvenated RBCs may be prepared and transfused within 24 hours or frozen for long-term storage.

DEGLYCEROLIZED REJUVENATED RED BLOOD CELLS is the form in which rejuvenated, cryopreserved red cells (Frozen Rejuvenated RBCs) are made available for infusion. For additional information, see sections on Rejuvenated RBCs and Deglycerolized RBCs above.

Plasma Components

Overview

Plasma is the fluid part of blood and can be derived from the separation of a whole blood collection or by apheresis collection. Important elements in plasma include albumin, coagulation factors, fibrinolytic proteins, immunoglobulin, and other proteins. Once plasma is collected, it can be maintained in the liquid state or stored frozen and subsequently thawed and kept in a liquid state. If Fresh Frozen Plasma (FFP) is thawed at 1 to 6 C, and the insoluble cryoprecipitate (see cryoprecipitated components) is removed by centrifugation, the supernatant plasma can be refrozen and labeled as Plasma Cryoprecipitate Reduced. Labile coagulation factor levels vary based upon ABO group, storage conditions, and/or further processing (see Tables 4 and 5).

Table 4. Coagulation Factor Activity in FFP and PF24 (whole blood) at the Time of Thaw and after 120 Hours of 1 to 6 C Storage (adapted from Table 1 in Scott EA, et al. Transfusion 2009;49:1584-91)

Analyte	Thaw, Mean ± SD (range) by Product		120 hr, Mean (range) by Product		% Change after 120 hr at 1 to 6 C	
	FFP (n=20)	PF24 (n=14)*	FFP (n=20)	PF24 (n=14)*	FFP	PF24
Factor II (IU/dL)	97 ± 10 (83-125)	97 ± 8 (80-113)	95 ± 10 (82-126)	96 ± 11 (74-120)	3 [†]	1
Factor V (U/dL)	85 ± 13 (63-104)	86 ± 16 (54-124)	67 ± 19 (17-92)	59 ± 22 (15-109)	21 [†]	31 [†]
Factor VII (IU/dL)	105 ± 25 (50-163)	89 ± 22 (54-145)	70 ± 18 (34-102)	77 ± 27 (50-159)	33 [†]	14 [†]
Factor VIII (IU/dL) [‡]	81 ± 19 (47-117)	66 ± 17 (30-100) [§]	43 ± 10 (27-60)	48 ± 12 (26-73)	47 [†]	28 [†]
Factor IX (IU/dL)	82 ± 13 (62-108)	88 ± 13 (70-105)	80 ± 12 (64-107)	84 ± 12 (65-99)	2	4 [†]
Factor X (IU/dL)	94 ± 10 (71-112)	94 ± 11 (72-112)	87 ± 11 (65-111)	91 ± 12 (67-114)	7 [†]	3 [†]
vWF:Ag (IU/dL) [‡]	98 ± 27 (57-156)	132 ± 41 (78-211)	97 ± 30 (48-150)	127 ± 40 (79-224)	1	4
vWF:RCo (IU/dL) [‡]	101 ± 26 (61-152)	123 ± 47 (58-238)	93 ± 30 (48-149)	102 ± 38 (50-191)	8 [†]	17 [†]
Fibrinogen (mg/dL)	280 ± 52 (223-455)	309 ± 70 (211-500)	278 ± 50 (223-455)	303 ± 50 (205-490)	1	2 [†]
Antithrombin (IU/dL)	97 ± 9 (85-118)	97 ± 11 (77-110)	100 ± 10 (85-131)	101 ± 14 (73-116)	3	4 [†]
Protein C (IU/dL)	107 ± 20 (74-148)	88 ± 16 (65-120) [§]	107 ± 19 (77-148)	89 ± 17 (65-115) [§]	0	2
Protein S (IU/dL)	97 ± 18 (61-123)	92 ± 18 (54-121)	90 ± 22 (52-134)	78 ± 19 (46-114) [§]	7 [†]	15 [†]

*N = 25 for Factor II, Factor V, Factor VIII, fibrinogen, vWF:RCo, and Protein S.

[†]p <0.05 when comparing mean activity at thaw to mean activity after 120 hours of 1 to 6 C storage.

[‡]Only results from group O products were used for statistical comparisons of Factor VIII, vWF:Ag, and vWF:RCo activities.

[§]p <0.05 compared with mean activity in FFP of the same age.

FFP = Fresh Frozen Plasma; PF24 = Plasma Frozen Within 24 Hours After Phlebotomy; SD = standard deviation; vWF:Ag = von Willebrand factor antigen; vWF:RCo = von Willebrand factor ristocetin cofactor.

Table 5. Statistically Significantly Different Coagulation Factor Activity in FFP and PF24RT24 (apheresis) after 24 Hours at 1 to 6 C Storage after Thawing [adapted from Tables 2 and 3 of the 102nd Meeting of the Blood Products Advisory Committee (May 16, 2012)]

Analyte	Sponsor A			Sponsor B		
	Mean ± SD (range) by Product		Mean Difference:	Mean ± SD (range) by Product		Mean Difference:
	FFP (n=52)	PF24RT24 (n=52)	PF24RT24 – FFP (95% CLs)	FFP (n=54)	PF24RT24 (n=54)	PF24RT24 – FFP (95% CLs)
Factor V (IU/dL)	101 ± 18 (52-138)	100 ± 17 (52-136)	-1.1 (-2.1, -0.1)*	90 ± 19 (35-136)	89 ± 18 (35-131)	-1.0 (-2.6, 0.6)
Factor VIII (IU/dL)	81 ± 25 (37-163)	73 ± 24 (36-157)	-7.3 (-9.4, -5.2)†	99 ± 32 (49-193)	86 ± 27 (40-156)	-13.2 (-16.0, -10.5)†
Protein S (IU/dL)	94 ± 20 (53-161)	83 ± 19 (48-145)	-10.6 (-12.7, -8.5)†	82 ± 18 (29-124)	73 ± 14 (47-109)	-9.0 (-11.7, -6.2)†

*p = <0.05.

†p = <0.0001.

CLs = confidence limits; FFP = Fresh Frozen Plasma; PF24RT24 = Plasma Frozen Within 24 Hours After Phlebotomy Held At Room Temperature Up to 24 Hours After Phlebotomy; SD = standard deviation.

Refer to the section on **Further Processing** for additional information on:

- Pathogen Reduction Technology and Components Available

Refer to the section on **Additional Testing** for additional information on:

- Identification of Low-Titer Anti-A and/or Anti-B Blood Products

Fresh Frozen Plasma

Description

FFP is prepared from a whole blood or apheresis collection and frozen at -18 C or colder within the time frame specified in the manufacturer's instructions for use of the blood collection, processing, and storage system. The anticoagulant solution used and the component volume are indicated on the label. On average, units contain 200 to 250 mL, but apheresis-derived units may contain as much as 400 to 600 mL. FFP contains plasma proteins, including all coagulation factors. FFP contains normal levels of the labile coagulation factors, Factors V and VIII.

FFP should be infused immediately after thawing or stored at 1 to 6 C. After 24 hours, the component must be discarded or, if collected in a functionally closed system, may be relabeled as Thawed Plasma Ω (see Thawed Plasma Ω).

Actions

FFP serves as a source of plasma proteins for patients who are deficient in or have defective plasma proteins.

Indications

FFP is indicated in the following conditions:

1. Management of preoperative or bleeding patients who require replacement of multiple plasma coagulation factors (eg, liver disease, DIC).
2. Patients undergoing massive transfusion who have clinically significant coagulation deficiencies.
3. Patients taking warfarin who are bleeding or need to undergo an invasive procedure before vitamin K could reverse the warfarin effector and need only transient reversal of warfarin effect.
4. Transfusion or plasma exchange in patients with thrombotic thrombocytopenic purpura (TTP).
5. Management of patients with selected coagulation factor deficiencies, congenital or acquired, for which no specific coagulation concentrates are available.
6. Management of patients with rare specific plasma protein deficiencies, such as Factor V deficiency, where recombinant products are unavailable.

Contraindications

1. Plasma is not indicated when coagulopathy can be corrected more effectively with specific therapy, such as vitamin K for urgent vitamin K antagonist (VKA) reversal, Cryoprecipitated AHF or Pathogen Reduced Cryoprecipitated Fibrinogen Complex for hypofibrinogenemia, or specific coagulation factor concentrates when available. Specific reversal agents should be used for non-VKA anticoagulants (eg, idarucizumab for dabigatran or andexanet for Factor Xa inhibitors such as rivaroxaban- and apixaban-related life-threatening bleeding).
2. Plasma transfusion is not indicated for volume expansion in the absence of a deficiency of plasma proteins. In such cases, blood volume can be safely and adequately replaced with other volume expanders.

Relative Contraindications

1. Plasma should not be transfused when certain deficiencies or coagulopathies can be corrected with more specific therapies or anticoagulant reversal agents.
2. Plasma should not be transfused to correct a minimally elevated international normalized ratio (INR) of 1.7 or less. An INR value between 1.5 and 1.7 represents at least 30% of coagulation factor levels, which should allow for normal hemostasis. Transfusion of a standard dose of plasma [$\sim 15\text{ mL/kg}$] to a patient with an INR of 1.7 or lower may not normalize the INR.

Dosage and Administration

Compatibility tests prior to transfusion are not necessary. Plasma must be ABO compatible with the recipient's red cells. Compatibility with Rh(D) is not necessary in plasma transfusion. The volume transfused depends on the clinical situation and patient size and may be guided by laboratory assays of coagulation function.

FFP must be thawed in a water bath at 30 to 37 C or in an FDA-cleared device for thawing plasma. If a water bath is used, thaw the component in a protective plastic overwrap using gentle agitation.

See Table 3 for pediatric dosage information.

Side Effects and Hazards

Do not use FFP if there is evidence of container breakage or of thawing during storage.

Hazards that pertain to all transfusion components, including FFP, are described in the earlier section on Side Effects and Hazards for Whole Blood and All Blood Components.

*Components Available***FRESH FROZEN PLASMA****APHERESIS FRESH FROZEN PLASMA****Plasma Frozen Within 24 Hours After Phlebotomy***Description*

Plasma Frozen Within 24 Hours After Phlebotomy (PF24) is prepared from a whole blood or apheresis collection. The anticoagulant solution used and the component volume are indicated on the label. On average, PF24 contains 200 to 250 mL, but apheresis-derived units may contain as much as 400 to 600 mL. This plasma component is a source of nonlabile plasma proteins.

Plasma proteins such as albumin; a disintegrin and metalloprotease with thrombospondin type 1 motifs 13 (ADAMTS13); fibrinogen; and Factors II, VII, IX, X, and XI remain at levels similar to FFP. Levels of Factor VIII and Protein C are reduced, and levels of Factor V and other labile plasma proteins are variable compared with FFP.

PF24 should be infused immediately after thawing or stored at 1 to 6 C. After 24 hours' storage, the component must be discarded or, if collected in a functionally closed system, may be relabeled as Thawed Plasma Ω (see Thawed Plasma Ω).

Actions

PF24 serves as a source of nonlabile plasma proteins for patients who are deficient in or have defective plasma proteins. Some coagulation factor levels may be lower than those of FFP, especially labile coagulation Factors V and VIII and Protein C.

Indications

For PF24 indications, see *Indications* in earlier section on Fresh Frozen Plasma.

Contraindications

For PF24 contraindications, see *Contraindications* and *Relative Contraindications* in earlier section on Fresh Frozen Plasma. In addition, this product is not indicated for treatment of deficiencies of labile coagulation factors, including Factors V and VIII, and Protein C.

Dosage and Administration

For PF24 dosage and administration, see *Dosage and Administration* in earlier section on Fresh Frozen Plasma.

Side Effects and Hazards

For PF24 side effects and hazards, see *Side Effects and Hazards* in earlier section on Fresh Frozen Plasma.

Components Available

PLASMA FROZEN WITHIN 24 HOURS AFTER PHLEBOTOMY is prepared from a whole blood collection and must be separated and placed at -18 C or colder within 24 hours from whole blood collection.

APHERESIS PLASMA FROZEN WITHIN 24 HOURS AFTER PHLEBOTOMY is prepared from apheresis and stored at 1 to 6 C within 8 hours of collection and frozen at -18 C or colder within 24 hours of collection.

Plasma Frozen Within 24 Hours After Phlebotomy Held At Room Temperature Up To 24 Hours After Phlebotomy*Description*

Plasma Frozen Within 24 Hours After Phlebotomy Held At Room Temperature Up To 24 Hours After Phlebotomy (PF24RT24) is prepared from whole blood or an apheresis collection. The product can be held at room temperature for up to 24 hours after collection and then frozen at -18 C or colder. The anticoagulant solution used and the component volume are indicated on the label. On average, PF24RT24 contains 200 to 250 mL, but apheresis-derived units may contain as much as 400 to 600 mL. This plasma component is a source of nonlabile plasma proteins.

Plasma proteins such as albumin; ADAMTS13; fibrinogen; and Factors II, VII, IX, X, and XI remain at levels similar to FFP. Levels of Factor V, Factor VIII, and Protein S are reduced, and levels of other labile plasma proteins are variable compared with FFP.

PF24RT24 should be infused immediately after thawing or stored at 1 to 6 C. After 24 hours, the component must be discarded or, if collected in a functionally closed system, may be relabeled as Thawed Plasma Ω (see Thawed Plasma Ω).

Actions

This plasma component serves as a source of nonlabile plasma proteins for patients who are deficient in or have defective plasma proteins. Some coagulation factor levels may be lower than those of FFP, especially labile coagulation Factors V and VIII and Protein S.

Indications

For PF24RT24 indications, see *Indications* in earlier section on Fresh Frozen Plasma.

Contraindications

For PF24RT24 contraindications, see *Contraindications* and *Relative Contraindications* in earlier section on Fresh Frozen Plasma. In addition, this product is not indicated for treatment of deficiencies of labile coagulation factors, including Factors V and VIII, and Protein S.

Dosage and Administration

For PF24RT24 dosage and administration, see *Dosage and Administration* in earlier section on Fresh Frozen Plasma.

Side Effects and Hazards

For PF24RT24 side effects and hazards, see *Side Effects and Hazards* in earlier section on Fresh Frozen Plasma.

Components Available

PLASMA FROZEN WITHIN 24 HOURS AFTER PHLEBOTOMY HELD AT ROOM TEMPERATURE UP TO 24 HOURS AFTER PHLEBOTOMY

APHERESIS PLASMA FROZEN WITHIN 24 HOURS AFTER PHLEBOTOMY HELD AT ROOM TEMPERATURE UP TO 24 HOURS AFTER PHLEBOTOMY

Plasma Cryoprecipitate Reduced

Description

Plasma Cryoprecipitate Reduced is prepared from whole-blood-derived or apheresis-collected plasma after thawing and centrifugation and removal of the cryoprecipitate. The remaining product is plasma that is reduced in fibrinogen, Factor VIII, Factor XIII, vWF, and cryoglobulin. This supernatant plasma must be refrozen within 24 hours of thawing at -18 C or colder. Proteins such as albumin, ADAMTS13, and Factors II, V, VII, IX, X, and XI remain at levels similar to the plasma product used to prepare the cryoprecipitate. High-molecular-weight forms of vWF (multimers) are significantly decreased during production; however, smaller multimers are retained.

Plasma Cryoprecipitate Reduced should be infused immediately after thawing or stored at 1 to 6 C. This product can be stored at 1 to 6 C for up to 4 days after the initial 24-hour postthaw period has elapsed, but must be relabeled as Thawed Plasma Cryoprecipitate Reduced Ω .

Actions

This component serves as a source for plasma proteins except for fibrinogen, Factor VIII, Factor XIII, and vWF.

Indications

Plasma Cryoprecipitate Reduced is used for transfusion or plasma exchange in patients with TTP. It may be used to provide clotting factors except fibrinogen, Factor VIII, Factor XIII, and vWF for transfusion support of patients with appropriate clinical indications when specific plasma concentrates and/or other plasma products are not available.

Contraindications

Plasma Cryoprecipitate Reduced is contraindicated for the repletion of coagulation factors known to be depleted in this product: fibrinogen, vWF, Factor VIII, and Factor XIII. This component should not be used as a substitute for FFP, PF24, PF24RT24, or Thawed Plasma.

Dosage and Administration

For Plasma Cryoprecipitate Reduced dosage and administration, see *Dosage and Administration* in earlier section on Fresh Frozen Plasma.

Side Effects and Hazards

For Plasma Cryoprecipitate Reduced side effects and hazards, see *Side Effects and Hazards* in earlier section on Fresh Frozen Plasma.

Components Available

PLASMA CRYOPRECIPITATE REDUCED
APHERESIS PLASMA CRYOPRECIPITATE REDUCED

Thawed Plasma Ω*Description*

Thawed Plasma is derived from FFP, PF24, or PF24RT24 prepared using aseptic techniques (functionally closed system). It is thawed at 30 to 37 C and maintained at 1 to 6 C for up to 4 days after the initial 24-hour postthaw period. The volume is indicated on the label. Thawed Plasma contains stable coagulation factors such as Factor II and fibrinogen in concentrations clinically similar to those of FFP, but variably reduced amounts of other factors (see Table 4).

Actions

This component serves as a source of nonlabile plasma proteins. Levels and activation state of coagulation proteins in Thawed Plasma are variable and change over time.

Indications

For Thawed Plasma indications, see *Indications* in earlier section on Fresh Frozen Plasma.

Contraindications

For Thawed Plasma contraindications, see *Contraindications* and *Relative Contraindications* in earlier section on Fresh Frozen Plasma. Do not use Thawed Plasma as the treatment for isolated coagulation factor or specific plasma protein deficiencies where other products are available with higher concentrations of the specific factor(s) or proteins.

Dosage and Administration

For Thawed Plasma dosage and administration, see *Dosage and Administration* in earlier section on Fresh Frozen Plasma.

Side Effects and Hazards

For Thawed Plasma side effects and hazards, see *Side Effects and Hazards* in earlier section on Fresh Frozen Plasma.

Components Available

THAWED PLASMA Ω

Thawed Plasma Cryoprecipitate Reduced Ω*Description*

Thawed Plasma Cryoprecipitate Reduced is derived from Plasma Cryoprecipitate Reduced. It is thawed at 30 to 37 C and maintained at 1 to 6 C for up to 4 days after the initial 24-hour postthaw period has elapsed. The volume is indicated on the label. Thawed Plasma Cryoprecipitate Reduced is deficient in fibrinogen, Factor VIII, Factor XIII, vWF, and cryoglobulin, and contains variable levels of albumin, ADAMTS13, and Factors II, V, VII, IX, X, and XI.

Actions

For Thawed Plasma Cryoprecipitate Reduced actions, see *Actions* in earlier section on Plasma Cryoprecipitate Reduced.

Indications

For Thawed Plasma Cryoprecipitate Reduced indications, see *Indications* in earlier section on Plasma Cryoprecipitate Reduced.

Contraindications

For Thawed Plasma Cryoprecipitate Reduced contraindications, see *Contraindications* in earlier section on Plasma Cryoprecipitate Reduced.

Dosage and Administration

For Thawed Plasma Cryoprecipitate Reduced dosage and administration, see *Dosage and Administration* in earlier section on Fresh Frozen Plasma.

Side Effects and Hazards

For Thawed Plasma Cryoprecipitate Reduced side effects and hazards, see *Side Effects and Hazards* in earlier section on Fresh Frozen Plasma.

*Components Available***THAWED PLASMA CRYOPRECIPITATE REDUCED Ω** **Liquid Plasma***Description*

Liquid Plasma is prepared from Whole Blood and stored at 1 to 6 C. Liquid Plasma expires 5 days from the end of the Whole Blood dating period.

The profile and activity of plasma proteins involved in coagulation of Liquid Plasma are not completely characterized. Levels and activation states of coagulation proteins in Liquid Plasma are dependent upon and change with time in contact with cells, as well as the conditions and duration of storage. This product contains viable lymphocytes that may cause graft-vs-host reactions in susceptible patients.

Actions

This component serves as a source of plasma proteins. Levels and activation states of coagulation proteins are variable and change over time.

Indications

Liquid Plasma is indicated for the initial treatment of patients who are undergoing massive transfusion because of life-threatening trauma/hemorrhages and who have clinically significant coagulation deficiencies.

Contraindications

For Liquid Plasma contraindications, see *Contraindications* in earlier section on Fresh Frozen Plasma. Do not use Liquid Plasma as the treatment for coagulation factor deficiencies where other products are available with higher factor concentrations.

Dosage and Administration

For Liquid Plasma dosage and administration, see *Dosage and Administration* in earlier section on Fresh Frozen Plasma.

Side Effects and Hazards

For Liquid Plasma side effects and hazards, see *Side Effects and Hazards* in earlier section on Fresh Frozen Plasma.

*Components Available***LIQUID PLASMA****Cryoprecipitated Antihemophilic Factor***Description*

Cryoprecipitated Antihemophilic Factor (AHF) is prepared by thawing frozen whole-blood-derived or apheresis plasma between 1 and 6 C and recovering the precipitate. The cold-insoluble precipitate is placed in the freezer at -18 C or colder within 1 hour after removal from the refrigerated centrifuge. Cryoprecipitated AHF contains fibrinogen, Factor VIII, Factor XIII, and vWF. Each unit of Cryoprecipitated AHF should contain ≥ 80 International Units (IU) of Factor VIII and ≥ 150 mg of fibrinogen in approximately 5 to 20 mL of plasma.

If the label indicates “Pooled Cryoprecipitated AHF,” several units of Cryoprecipitated AHF have been pooled. The volume of the pool is indicated on the label and, if used, the volume of 0.9% Sodium Chloride, Injection USP may be separately listed. To determine the minimum potency of this component, assume 80 IU of Factor VIII and 150 mg of fibrinogen for each unit of Cryoprecipitated AHF indicated on the label.

Actions

Cryoprecipitate serves as a source of fibrinogen, Factor VIII, Factor XIII, and vWF.

Indications

This component is used in the control of bleeding associated with fibrinogen deficiency, and when recombinant and/or virally inactivated preparations of fibrinogen, Factor VIII, Factor XIII, or vWF are not readily available. It is also indicated as second-line therapy for von Willebrand disease (vWD) and hemophilia A (Factor VIII deficiency). Coagulation factor preparations other than Cryoprecipitated AHF are preferred for management of vWD, Factor VIII deficiency, and Factor XIII deficiency. Every effort must be made to obtain preferred factor concentrates for hemophilia A patients before resorting to the use of Cryoprecipitated AHF. Use of this component may be considered for control of uremic bleeding after other modalities have failed.

Contraindications

Do not use this component unless results of laboratory studies indicate a specific hemostatic defect for which this product is indicated. Cryoprecipitated AHF should not be used if virus-inactivated specific factor concentrates or recombinant factor preparations are available for management of patients with vWD, hemophilia A, or Factor XIII deficiency.

Dosage and Administration

Compatibility testing is unnecessary. ABO-compatible Cryoprecipitated AHF may be preferred in neonates. Rh type need not be considered when using this component.

The frozen component is thawed in a protective plastic overwrap in a water bath at 30 to 37 C or FDA-cleared device up to 15 minutes (thawing time may be extended if product is pooled before freezing). This component should not be given if there is evidence of container breakage or of thawing during storage. Do not refreeze after thawing. Thawed Cryoprecipitated AHF should be kept at room temperature and transfused as soon as possible after thawing, within 6 hours if it is a single unit (from an individual donor, or products pooled before freezing or prior to administration using an FDA-cleared sterile connecting device), and within 4 hours after entering the container (eg, to attach an administration set or to pool) without using an FDA-cleared sterile connecting device.

Cryoprecipitated AHF may be transfused as individual units or pooled. For pooling, the precipitate in one or more concentrates should be mixed well with 10 to 15 mL of diluent to ensure complete removal of all material from the container. The preferred diluent is 0.9% Sodium Chloride, Injection USP. Serial use of each bag's contents to resuspend the precipitate into subsequent bags may be used to efficiently pool cryoprecipitate into a single bag.

The recovery of transfused fibrinogen is 50% to 60%. When used to correct hypofibrinogenemia, Cryoprecipitated AHF may be dosed at one bag per 7 to 10 kg body weight to raise plasma fibrinogen by approximately 50 to 75 mg/dL. Thrombosis alters fibrinogen kinetics; therefore, patients receiving cryoprecipitate as fibrinogen replacement in conditions associated with increased fibrinogen turnover should be monitored with fibrinogen or viscoelastic testing assays. For treatment of bleeding in patients with hemophilia A when Factor VIII concentrates are not available, rapid infusion of a loading dose expected to produce the desired level of Factor VIII is usually followed by a smaller maintenance dose every 8 to 12 hours. To maintain hemostasis after surgery, a regimen of therapy for 10 days or longer may be required. If circulating antibodies to Factor VIII are present, the use of larger doses, activated concentrates, porcine-derived concentrates, or other special measures may be indicated. To calculate cryoprecipitate dosage as a source of Factor VIII, the following formula is helpful: $\text{Number of bags} = (\text{Desired increase in Factor VIII level in } \% \times 40 \times \text{body weight in kg}) / \text{average units of Factor VIII per bag}$. Optimal patient management requires that the Cryoprecipitated AHF treatment responses of Factor VIII-deficient recipients be monitored with periodic plasma Factor VIII assays.

For treatment of vWD, smaller amounts of Cryoprecipitated AHF will usually be adequate. Because the vWF content of Cryoprecipitated AHF is not usually known, an empiric dose of 1 bag per 10 kg of body weight has been recommended. Patients receiving this treatment should be monitored by appropriate laboratory studies to determine the frequency of Cryoprecipitated AHF administration.

See Table 3 for pediatric dosage information.

Side Effects and Hazards

Hazards that pertain to all transfusion components are described in the earlier section on Side Effects and Hazards for Whole Blood and All Blood Components.

If a large volume of ABO-incompatible Cryoprecipitated AHF is used, the recipient may develop a positive DAT.

Components Available

CRYOPRECIPITATED AHF

APHERESIS CRYOPRECIPITATED AHF

POOLED CRYOPRECIPITATED AHF

Platelet Components

Overview

Platelet transfusions are administered to treat patients with thrombocytopenia, dysfunctional platelet disorders, or active platelet-related bleeding, or administered prophylactically to patients at serious risk of bleeding. Conventional platelet components are stored at room temperature (20-24 C), in plasma or platelet additive solution (PAS), and include platelets manufactured by automated methods (apheresis platelets), as well as whole-blood-derived (WBD) single and pooled (prestorage and poststorage) platelet components.

Refer to the section on **Further Processing** for additional information on:

- Pathogen Reduction Technology and Components Available
- Leukocyte Reduction
- Irradiation
- Washing and Volume Reduction

Refer to the section on **Additional Testing** for additional information on:

- Identification of CMV-Seronegative Blood
- Identification of Low-Titer Anti-A and/or Anti-B Blood Products

Refer to the **June 2023 FDA Guidance, *Alternative Procedures for the Manufacture of Cold-Stored Platelets Intended for the Treatment of Active Bleeding when Conventional Platelets Are Not Available or Their Use Is Not Practical***, for information on the use of Cold-Stored Platelets.

Description

Platelets for transfusion are manufactured using automated collection by apheresis (“Apheresis Platelets”) or from whole blood collections (“WBD Platelets”). One unit of WBD Platelets typically contains $\geq 5.5 \times 10^{10}$ platelets suspended in 40 to 70 mL of plasma. WBD Platelets may be transfused as single units or as a pool. WBD Platelets may be pooled prestorage using a closed system or poststorage using an open system. A pool of approximately 6 units of WBD Platelets is considered the therapeutic equivalent of 1 unit of Apheresis Platelets, which usually contains $\geq 3.0 \times 10^{11}$ platelets.

Platelet components may contain a varying number of leukocytes depending upon the manufacturing method. Some units may contain more than the trace amounts of red cells usually present and will appear pink to salmon in color. This occurs more frequently with WBD platelets than with apheresis platelets. Platelet products stored in 100% plasma at room temperatures also contain variable levels of stable coagulation factors.

FDA has provided recommendations for bacterial risk control strategies in the December 2020 Final Guidance, *Bacterial Risk Control Strategies for Blood Collection Establishments and Transfusion Services to Enhance the Safety and Availability of Platelets for Transfusion*. To control the risk of bacterial contamination, platelets are pathogen reduced, tested for bacteria, or stored at 1 to 6 C using procedures found acceptable by FDA and FDA-cleared or -approved devices according to the manufacturer’s instructions for use.

Conventional platelet products are stored at room temperature, 20-24 C, with continuous gentle agitation. Based on the strategy used, conventional platelets may have either a 5-, 6-, or 7-day expiration. Note that certain testing strategies may require secondary testing prior to transfusion.

For more information on bacterial contamination risk refer to *Side Effects and Hazards* section below and FDA Guidance titled *Bacterial Risk Control Strategies for Blood Collection Establishments and Transfusion Services to Enhance the Safety and Availability of Platelets for Transfusion*.

Actions

Platelets are essential for normal hemostasis. Complex reactions occur between platelets, vWF, collagen in the walls of disturbed vasculature, phospholipids, and soluble coagulation factors, including thrombin. These changes induce platelet adherence to vessel walls and platelet activation, which leads to platelet aggregation and formation of a primary hemostatic plug. The therapeutic goal of platelet transfusion is to provide adequate numbers of normally functioning platelets for the prevention or cessation of bleeding.

Indications

Platelet transfusions may be given to patients with thrombocytopenia, dysfunctional platelet disorders (congenital, metabolic, or medication-induced), or active platelet-related bleeding, or patients at serious risk of bleeding (ie, prophylactic use). Patients with the following medical conditions may require platelet transfusion: leukemia, myelodysplasia, aplastic anemia, solid tumors, congenital or acquired platelet dysfunction, and central nervous system trauma. Patients undergoing extracorporeal membrane oxygenation or cardiopulmonary bypass may also need platelet transfusion, and platelets are frequently indicated in massive transfusion protocols. Thrombocytopenia is unlikely to be the solitary cause of bleeding in patients with platelet counts of at least 50,000/microliter (μL). Higher transfusion thresholds may be appropriate for patients with central nervous system bleeds or platelet dysfunction. For the clinically stable patient with an intact vascular system and normal platelet function, prophylactic platelet transfusions may be appropriate at <5000 to $10,000/\mu\text{L}$.

Prophylactic platelet transfusion may not be of therapeutic benefit when thrombocytopenia is related to destruction of circulating platelets secondary to autoimmune disorders [eg, immune thrombocytopenic purpura (ITP)]; however, transfusion may be indicated for active bleeding in these patients. Platelets/pooled Platelets Leukocytes Reduced or Apheresis Platelets Leukocytes Reduced are indicated to decrease the frequency of recurrent febrile nonhemolytic transfusion reaction, HLA alloimmunization, and transfusion-transmitted CMV infection (see sections on **Further Processing** and **Additional Testing**).

Contraindications

Do not use this component if bleeding is unrelated to decreased numbers of, or abnormally functioning, platelets. Platelets should not be transfused when the platelet count is greater than $100,000/\mu\text{L}$ unless there is documented or suspected abnormal function. Prophylactic transfusion is generally not indicated in nonbleeding patients on antiplatelet medications, or when platelet dysfunction is extrinsic to the platelet, such as in uremia, certain types of vWD, and hyperglobulinemia. Patients with congenital surface glycoprotein defects should be transfused conservatively to reduce the possibility for alloimmunization to the missing protein(s).

Do not use in patients with activation or autoimmune destruction of endogenous platelets, such as in heparin-induced thrombocytopenia (HIT), TTP, or ITP, unless the patient has a life-threatening hemorrhage.

Dosage and Administration

Compatibility testing is not necessary in routine platelet transfusion. Donor plasma should be ABO compatible with the recipient's red cells when this component is to be transfused to pediatric patients with small blood volumes. The number of platelet units to be administered depends on the clinical situation of each patient. An apheresis platelet unit, transfused to an average-sized relatively healthy recipient, would be expected to result in a 1-hour posttransfusion increase in platelet count of approximately 30,000 to $60,000/\mu\text{L}$. One unit of WBD platelets would be expected to increase the platelet count of a 70-kg adult by 5000 to $10,000/\mu\text{L}$ and increase the count of an 18-kg child by $20,000/\mu\text{L}$. The therapeutic adult dose is 1 unit of apheresis platelets or 6 units of WBD platelets, either of which usually contains $\geq 3.0 \times 10^{11}$ platelets. For prophylaxis, this dose may need to be repeated in 1 to 3 days because of the short life span of transfused platelets (3 to 4 days). Platelet components must be examined for abnormal appearance before administration. Units with excessive aggregates should not be administered. The transfusion of a platelet unit, using a standard administration set, may proceed as quickly as tolerated, but must take less than 4 hours after entering the container.

The corrected count increment (CCI) for conventional platelets transfused to nonbleeding patients is a calculated measure of patient response to platelet transfusion and is not directly correlated with bleeding risk. CCI adjusts for the number of platelets infused and the size of the recipient, based upon body surface area (BSA):

$$\text{CCI} = (\text{postcount} - \text{precount}) \times \text{BSA} / \text{platelets transfused}$$

where *postcount* and *precount* are platelet counts ($/\mu\text{L}$) after and before transfusion, respectively; *BSA* is patient BSA (meter^2); and *platelets transfused* is the number of administered platelets ($\times 10^{11}$). The CCI is usually determined 10 to 60 minutes after transfusion is completed. For example:

A patient with acute myelogenous leukemia with a nomogram-derived BSA of 1.40 m² is transfused with a unit of Apheresis Platelets (a platelet dose of 4.5×10^{11}). The pretransfusion platelet count is 2000/ μ L. The patient's platelet count from a sample of blood collected 15 minutes after platelet transfusion is 29,000/ μ L. The CCI is calculated as $(29,000 - 2000) \times 1.4 / 4.5 = 8400/\mu\text{L per } 10^{11} \text{ per m}^2$.

In an afebrile, nonbleeding patient, the CCI is typically greater than 7500 at 10 minutes to 1 hour after transfusion and remains above 4500 at 24 hours for conventional platelets that are not pathogen reduced. A lower CCI may be expected following transfusion with platelet components that have been further manufactured (pathogen reduced, irradiated, or washed) or in patients that have been multiply transfused. Both immune and nonimmune mechanisms of platelet destruction may contribute to reduced platelet recovery and lower CCIs. Along with supportive serologic test results, a CCI of less than 5000 at 10 minutes to 1 hour after transfusion may indicate an immune-mediated refractory state to platelet therapy (refer to Platelet Alloimmunization, below). With nonimmune mechanisms, platelet recovery within 1 hour may be adequate, although survival at 24 hours is reduced.

See Table 3 for pediatric dosage information.

Side Effects and Hazards

Hazards that pertain to all transfusion components are described in the section on Side Effects and Hazards for Whole Blood and All Blood Components. Listed below is **additional** information on hazards or hazards more often associated with platelet components.

1. **Bacterial Contamination:** Conventional room-temperature-stored platelets are associated with a higher risk of sepsis and related fatality than any other transfusable blood component. Thus, conventional platelet products either undergo bacterial detection testing as described above or undergo treatment using pathogen reduction technology approved by FDA. Although methods to limit and detect bacterial contamination have been implemented for platelet components, risk of bacterial contamination remains a hazard of platelet transfusion, and platelets remain the most likely blood components to be contaminated with bacteria. Gram-positive skin flora are the most commonly recovered bacteria. Symptoms may include, but are not limited to, high fever (≥ 2.0 C or ≥ 3.5 F increase in temperature), severe chills, hypotension, or circulatory collapse during or immediately after transfusion. In some instances, symptoms, especially when associated with contamination by gram-positive organisms, may be delayed for several hours following transfusion. Prompt management should include broad-spectrum antibiotic therapy along with cultures from the patient, suspected blood component(s), and administration set. Consider Gram stain, culture, or other rapid detection method of the suspected contaminated unit(s) whenever possible.
2. **Platelet Alloimmunization:** Platelets bear a variety of antigens, including Class I HLA and platelet-specific antigens. In the setting of platelet transfusion, patients may develop Class I HLA and/or human platelet antigen (HPA) antibodies, potentially leading to refractoriness to transfused platelets. When platelets are transfused to a patient with an antibody specific for an expressed antigen, the survival time of the transfused platelets may be markedly shortened. Medication should be considered as a cause of immune or nonimmune thrombocytopenia. Nonimmune events may also contribute to reduced platelet survival. It may be possible to distinguish between immune and nonimmune platelet refractoriness by assessing platelet recovery soon after infusion (ie, a 10- to 60-minute CCI). In immune refractory states secondary to serologic incompatibility, there is poor recovery in the early postinfusion interval, resulting in a CCI < 7500 . In nonimmune mechanisms (eg, splenomegaly, sepsis, fever, intravascular devices, and DIC), platelet recovery within 1 hour of infusion may be adequate, while longer-term survival (ie, 24-hour survival) is reduced. Serologic tests may confirm the presence of alloimmunization. Laboratory tests (HLA/HPA typing and antibody identification, or a platelet crossmatch) may also be helpful in selecting platelets with acceptable survival.
3. **Red Cell Alloimmunization:** Immunization to red cell antigens may occur because of the presence of residual red cells in platelets. Red cell compatibility testing is necessary only if the platelet component is prepared by a method that results in the component containing 2 mL or more of red cells, making the unit appear pink to salmon in color. This occurs more frequently with WBD platelets than apheresis platelets. Rh(D)-positive platelet transfusions to Rh(D)-negative individuals are common. The risk of Rh(D) alloimmunization is higher with WBD platelets and is very low with apheresis platelets. Providers may consider the use of Rh Immune Globulin to mitigate this risk in select patient populations.
4. **Hemolysis:** Platelet components that are not ABO identical with the recipient's blood group may contain incompatible plasma and, when transfused, may cause a positive DAT and possibly hemolysis. Platelet

transfusions from ABO-incompatible donors with isohemagglutinins (anti-A or anti-B) may cause acute hemolytic reactions in susceptible patients.

Components Available

This information is divided into sections by component type:

- WBD platelets
- Apheresis platelets

Whole-Blood-Derived Platelet Components:

PLATELETS are a concentrate of platelets separated from a single unit of Whole Blood, also referred to as WBD. One unit of Platelets should contain $\geq 5.5 \times 10^{10}$ platelets suspended in 40 to 70 mL of plasma. This component is usually provided as a pool. See below.

POOLED PLATELETS may be prepared using aseptic technique as an open or closed system. The number of units of Platelets in the pool will be indicated on the label. To determine the minimum potency of this component, assume 5.5×10^{10} platelets per unit of Platelets indicated on the label. See the label for the approximate volume.

PLATELETS LEUKOCYTES REDUCED may be prepared using an open or closed system. One unit of Platelets Leukocytes Reduced should contain $\geq 5.5 \times 10^{10}$ platelets and $< 8.3 \times 10^5$ leukocytes. This component is usually provided as a pool. See below.

POOLED PLATELETS LEUKOCYTES REDUCED may be prepared using aseptic technique as an open or closed system by pooling and filtering Platelets or pooling Platelets Leukocytes Reduced. The number of units in the pool will be indicated on the label. To determine the minimum potency of this component, assume 5.5×10^{10} platelets per unit of Platelets Leukocytes Reduced indicated on the label and $< 5 \times 10^6$ leukocytes in the pool. See the label for the approximate volume.

Apheresis Platelet Components:

APHERESIS PLATELETS are a therapeutic adult dose of platelets suspended in plasma collected from a single donor via an apheresis device. Apheresis Platelets should contain $\geq 3.0 \times 10^{11}$ platelets; 1 unit of Apheresis Platelets may be equivalent to 6 units of WBD Platelets. The product volume is variable and indicated on the label. The number of leukocytes contained in this component varies depending upon the blood cell separator and protocol used for collection. Apheresis Platelets are supplied in one or more connected bags to improve platelet viability during storage by providing more surface area for gas exchange. Anticoagulant citrate-dextrose solution A is the anticoagulant solution currently used for the collection and preservation of Apheresis Platelets.

APHERESIS PLATELETS LEUKOCYTES REDUCED can be leukocyte reduced during the collection process or may be prepared by further processing using leukocyte reduction filters. Apheresis Platelets Leukocytes Reduced should contain $\geq 3.0 \times 10^{11}$ platelets and $< 5.0 \times 10^6$ leukocytes. When Apheresis Platelets Leukocytes Reduced are prepared during further processing, these may be labeled Apheresis Platelets Leukocytes Reduced provided the requirement for residual leukocyte count is met and the platelet recovery is at least 85% of the pre-filtration content. The volume, anticoagulant-preservative, and storage conditions for Apheresis Platelets Leukocytes Reduced are the same as those for Apheresis Platelets.

APHERESIS PLATELETS PLATELET ADDITIVE SOLUTION ADDED LEUKOCYTES

REDUCED are platelets collected by apheresis and suspended in variable amounts of plasma and an FDA-approved PAS. See Table 6. One unit of platelets should contain $\geq 3.0 \times 10^{11}$ platelets and $< 5.0 \times 10^6$ leukocytes. The volume in the product is variable and indicated on the label. Plasma proteins, including coagulation factors present in the plasma, are diluted in proportion to the PAS added.

Refer to the section on **Further Processing** for information on pathogen-reduced platelet components.

Granulocyte Components

Apheresis Granulocytes Ω

Description

Apheresis Granulocytes contain numerous leukocytes and platelets, as well as 20 to 50 mL of red cells. The number of granulocytes in each concentrate is usually $> 1.0 \times 10^{10}$. The number of platelets varies in each prod-

Table 6. Contents of Platelet Additive Solutions

Additive Solution (mg/100 mL)	Sodium Chloride	Sodium Citrate	Sodium Gluconate	Sodium Acetate	Dibasic Sodium Phosphate	Monobasic Sodium Phosphate	Monobasic Potassium Phosphate	Potassium Chloride	Magnesium Chloride	Shelf Life (days)
PAS-C (Intersol)	452	318 (dihydrate)		442 (trihydrate)	305 (anhydrous)	93 (monohydrate)				5
PAS-F (Isoplate)	530		500	370 (trihydrate)	12 (heptahydrate)		0.82	37	30 (hexahydrate)	5

PAS = platelet additive solution.

uct. Various modalities may be used to improve granulocyte collection, including donor administration of granulocyte colony-stimulating factor and/or corticosteroids. The final volume of the product is 200 to 300 mL, including anticoagulant and plasma as indicated on the label.

Red cell sedimenting agents approved by FDA, such as hydroxyethyl starch (HES), are typically used in the collection of granulocytes. Residual sedimenting agents will be present in the final component and are described on the label. Apheresis Granulocytes should be administered as soon after collection as possible because of well-documented deterioration of granulocyte function during short-term storage. If stored, maintain at 20 to 24 C without agitation, for no more than 24 hours.

Actions

Granulocytes migrate toward, phagocytize, and kill bacteria and fungi. A quantitative relationship exists between the level of circulating granulocytes and the prevalence of bacterial and fungal infection in neutropenic patients. The ultimate goal is to provide the patient with the ability to fight infection. The infusion of a granulocyte component may not be associated with a significant increase in the patient's granulocyte count and is dependent on multiple factors, including the patient's clinical condition.

Indications

Granulocyte transfusion therapy is controversial. Apheresis Granulocytes are typically used in the treatment of patients with documented infections (bacterial and fungal) unresponsive to antimicrobial therapy in the setting of neutropenia [absolute granulocyte count $<0.5 \times 10^9/L$ ($<500/\mu L$)] with expected eventual marrow recovery. A trial of broad-spectrum antimicrobial agents should be used before granulocyte transfusion therapy is initiated. If the intended recipient is CMV seronegative and severely immunosuppressed (eg, a marrow transplant recipient), serious consideration should be given before administration of CMV-seropositive granulocytes. In addition to neutropenic patients, patients with hereditary neutrophil function defects (such as chronic granulomatous disease) may be candidates for granulocyte transfusion therapy.

Contraindications

Prophylactic use of granulocytes in noninfected patients is not routinely recommended. Patients with HLA and/or human neutrophil antigen (HNA) antibodies may not derive full benefit from granulocyte transfusion and may have a higher risk of pulmonary reactions. Antigen-matched or HLA-matched components, if available, may be considered in these patients.

Dosage and Administration

Transfuse as soon as possible and within 24 hours of collection. Due to the short shelf life of the product, these components may be released for emergency use prior to completion of infectious disease testing. A standard blood infusion set is to be used for the administration of Apheresis Granulocytes. Do not administer using leukocyte reduction filters. Depth-type microaggregate filters and leukocyte reduction filters remove granulocytes.

The red cells in Apheresis Granulocytes must be ABO compatible. Serologic compatibility between recipient and donor must be established before any red-cell-containing component is transfused. This may be accomplished by performing ABO/Rh typing, antibody screening, and crossmatching by serologic technique or use of a computer crossmatch.

Once granulocyte transfusion therapy is initiated, support should continue at least daily until infection is cured, defervescence occurs, the absolute granulocyte count returns to at least $0.5 \times 10^9/L$ ($500/\mu L$), or the physician in charge decides to halt the therapy.

Because most patients receiving these products are severely immunosuppressed and may be at risk for TA-GVHD, Apheresis Granulocytes must be irradiated (see sections on Further Processing and Additional Testing).

See Table 3 for pediatric dosage information.

Side Effects and Hazards

Hazards that pertain to all transfusion components are described in the section on Side Effects and Hazards for Whole Blood and All Blood Components. Listed below are hazards that apply specifically to Apheresis Granulocytes.

1. **Febriile Nonhemolytic Reactions:** These reactions are frequently noted in patients receiving granulocyte transfusions. Fever and chills in patients receiving granulocyte components may be avoided or mitigated by slow administration and recipient premedication.
2. **Allergic Reactions:** Allergic reactions to HES and other red-cell-sedimenting solutions may occur during granulocyte transfusion.

3. **Pulmonary Reactions:** Granulocyte transfusion can cause worsening of pulmonary function in patients with pneumonia, and rarely severe pulmonary reactions, especially in patients receiving concomitant amphotericin B. Patients who have pulmonary reactions should be tested for HLA and HNA antibodies.
4. **Alloimmunization:** Immunization to HLA antigens frequently occurs with granulocyte transfusion and can cause refractoriness to subsequent granulocyte or platelet transfusions.

Components Available

APHERESIS GRANULOCYTES Ω

Further Processing

This section addresses further processing of previously described blood components. The processes described in this section are pathogen reduction technology, leukocyte reduction, irradiation, washing, and volume reduction. A component may undergo one or more of these processes.

Pathogen Reduction Technology

Description

Pathogen reduction is an ex-vivo process intended to reduce the risk of certain transfusion-transmitted infections, including sepsis, and may also be used as an alternative to irradiation to prevent TA-GVHD if the pathogen reduction technology has been shown to inactivate residual lymphocytes. There is no pathogen inactivation process that has been shown to eliminate all pathogens; for example, hepatitis A (HAV), hepatitis E (HEV), human parvovirus B19, poliovirus, and *Bacillus cereus* spores have shown resistance to some processes. Apheresis Platelets Leukocytes Reduced Psoralen-Treated should contain $\geq 3.0 \times 10^{11}$ platelets and $< 5.0 \times 10^6$ leukocytes.

A current pathogen reduction procedure uses a chemical photosensitizer that is added to the plasma or platelet product and then transferred into a container that is placed inside an illumination device for ultraviolet A light (UVA) treatment. Unreacted photosensitizer and free photoproducts are subsequently removed with a compound adsorption device.

Products currently approved by FDA for pathogen reduction technology include apheresis platelets and whole-blood-derived (WBD) plasma or apheresis plasma. Pathogen-reduced plasma may be further manufactured, using a system approved by FDA for this purpose, into Pathogen Reduced Cryoprecipitated Fibrinogen Complex (PRCFC) and Pathogen Reduced Plasma Cryoprecipitate Reduced (PRPCR). Pathogen reduction technology may apply to other products in the future.

Consistent with the **Notice to All Users** section, refer to the manufacturer's instructions for use of components prepared using a pathogen reduction device for all components listed in this section.

- **Refer to the Platelet Components section or the Plasma Components section** for the corresponding *Description, Actions, Indications, Contraindications, Relative Contraindications, Dosage and Administration, and Side Effects and Hazards* as applicable to pathogen-reduced platelet components, and frozen and thawed pathogen-reduced plasma components.
- NOTE: *Additional Contraindications* for pathogen-reduced platelet and plasma components include:
 1. Contraindicated for preparation of pathogen-reduced components intended for patients with a history of hypersensitivity reaction to amotosalen or other psoralens.
 2. Contraindicated for preparation of pathogen-reduced components intended for neonatal patients treated with phototherapy devices that emit a peak energy wavelength less than 425 nanometers (nm) or have a lower bound of the emission bandwidth < 375 nm, due to the potential for erythema resulting from interaction between ultraviolet light and amotosalen.
- NOTE: Additional *Warnings and Precautions* for pathogen-reduced platelet and plasma components include:

Plasma Components:

Amotosalen-treated plasma may cause the following adverse reaction:

Cardiac Events:

In a randomized controlled trial of therapeutic plasma exchange (TPE) for TTP, five patients treated with INTERCEPT Blood System processed plasma and none with conventional plasma had adverse events in the cardiac system organ class (SOC) reported. These events included angina pectoris (n=3), cardiac arrest (n=1), bradycardia (n=1), tachycardia (n=1), and sinus arrhythmia (n=1). None of these events resulted in

documented myocardial infarction or death. Monitor patients for signs and symptoms of cardiac events during TPE for TTP.

Components Available

APHERESIS PLATELETS LEUKOCYTES REDUCED PSORALEN-TREATED
APHERESIS PLATELETS PLATELET ADDITIVE SOLUTION ADDED LEUKOCYTES
REDUCED PSORALEN-TREATED
APHERESIS FRESH FROZEN PLASMA PSORALEN-TREATED
POOLED FRESH FROZEN PLASMA PSORALEN-TREATED
POOLED PLASMA FROZEN WITHIN 24 HOURS AFTER PHLEBOTOMY
PSORALEN-TREATED
APHERESIS PLASMA CRYOPRECIPITATE REDUCED PSORALEN-TREATED
POOLED PLASMA CRYOPRECIPITATE REDUCED PSORALEN-TREATED
THAWED APHERESIS PLASMA PSORALEN-TREATED
THAWED POOLED PLASMA PSORALEN-TREATED
THAWED PLASMA CRYOPRECIPITATE REDUCED PSORALEN-TREATED

Pathogen Reduced Cryoprecipitated Fibrinogen Complex

Description

PRCFC is prepared from plasma that has been processed with an FDA-approved pathogen reduction device. The PRCFC process includes thawing pathogen-reduced plasma between 1 and 6 C and recovering the precipitate. The cold-insoluble precipitate is placed in the freezer at -18 C or colder.

Actions

PRCFC serves as an enriched source of fibrinogen, Factor XIII, vWF, and other constituents. The 5-day post-thaw shelf life of PRCFC is based on retention of critical functional activities that have shown a high level of correlation with therapeutic efficacy and the reduced pathogen risk associated with pathogen inactivation.

PRCFC is not intended to be used for replacement of Factor VIII.

Indications

PRCFC is indicated for:

1. Treatment and control of bleeding, including massive hemorrhage, associated with fibrinogen deficiency.
2. Control of bleeding when recombinant and/or specific virally inactivated preparations of Factor XIII or vWF are not available.
3. Second-line therapy for vWD.
4. Control of uremic bleeding after other treatment modalities have failed.

Limitations of Use: PRCFC should not be used for replacement of Factor VIII.

Contraindications

1. Contraindicated for preparation of blood components intended for patients with a history of hypersensitivity reaction to amotosalen or other psoralens.
2. Contraindicated for preparation of blood components intended for neonatal patients treated with phototherapy devices that emit a peak energy wavelength less than 425 nm or have a lower bound of the emission bandwidth <375 nm, due to the potential for erythema resulting from interaction between ultraviolet light and amotosalen.

Warnings and Precautions

1. For management of patients with vWD or Factor XIII deficiency, PRCFC should not be used if recombinant or specific virally inactivated factor preparations are available. In emergent situations, if recombinant or specific virally inactivated factor preparations are not available, PRCFC may be administered.

Dosage and Administration

1. Compatibility testing is not required. ABO-compatible PRCFC may be preferred in neonates. Rh type need not be considered when using this product.

2. Thaw according to institutional procedures and manufacturer's instructions for use of PRCFC. If using a water bath for thawing PRCFC, place in liquid-impermeable plastic overwrap. Do not allow product to contact water. Do not refreeze postthaw.
3. Do not administer PRCFC if there is evidence of container breakage or of thawing during frozen storage.
4. If PRCFC is pooled or aliquoted postthaw without using an FDA-cleared sterile connection device, transfuse within 4 hours of pooling or aliquoting.

PRCFC may be transfused from a single container or multiple containers. For in-hospital pooling, the precipitate in one or more containers may be mixed well with 10 to 15 mL of diluent to allow complete removal of all material from the container. The preferred diluent is 0.9% Sodium Chloride, Injection (USP). Serial use of each container's contents to resuspend the precipitate into subsequent containers may be used to efficiently pool PRCFC into a single container.

Thrombosis alters fibrinogen kinetics; therefore, patients receiving PRCFC as fibrinogen replacement in conditions associated with increased fibrinogen turnover should be monitored with fibrinogen assays.

When used to correct hypofibrinogenemia, PRCFC may be dosed based on the clinical presentation and expected fibrinogen content of the product. For example, a unit of PRCFC prepared from 2 WBD plasma units will contain about 740 ± 166 mg fibrinogen immediately postthaw, and 686 ± 165 mg fibrinogen after 120 hours.

Side Effects and Hazards

Hazards that pertain to all transfusion components are described in the earlier section on Side Effects and Hazards for Whole Blood and All Blood Components.

Components Available

**POOLED FIBRINOGEN COMPLEX CRYOPRECIPITATED PSORALEN-TREATED
APHERESIS FIBRINOGEN COMPLEX CRYOPRECIPITATED PSORALEN-TREATED
APHERESIS POOLED FIBRINOGEN COMPLEX CRYOPRECIPITATED PSORALEN-TREATED**

Leukocyte Reduction

Description

A unit of whole blood generally contains ≥ 1 to 10×10^9 white cells. Leukocyte reduction will decrease the cellular content and volume of blood according to characteristics of the technology used. RBCs Leukocytes Reduced, Apheresis RBCs Leukocytes Reduced, Apheresis Platelets Leukocytes Reduced, and Pooled Platelets Leukocytes Reduced must have $< 5.0 \times 10^6$ residual leukocytes per unit. Platelets Leukocytes Reduced (single unit WBD) must have $< 8.3 \times 10^5$ residual leukocytes per unit. Leukocyte reduction may be performed using in-process collection methods. Leukocyte reduction may be performed using additional postcollection processing steps to permit labeling as a leukocyte-reduced component: 1) soon after collection (prestorage), 2) after varying periods of storage in the laboratory, or 3) at the bedside as directed by the manufacturer's instructions. The methods used by the laboratory for leukocyte reduction are subject to quality control testing; leukocyte-reduced components prepared at the bedside are not routinely subjected to quality control testing. Leukocyte reduction technologies variably remove other cellular elements in addition to white cells. Washing is not a substitute for leukocyte reduction. Leukocyte reduction is not a substitute for irradiation.

Indications

Leukocyte-reduced components are indicated to decrease the frequency of recurrent febrile nonhemolytic transfusion reaction. They have also been shown to reduce the risk of transfusion-transmitted CMV and to reduce the incidence of HLA alloimmunization.

Contraindications

Leukocyte-reduced components do not prevent TA-GVHD.

Leukocyte reduction filters are not to be used in the administration of Apheresis Granulocytes.

Side Effects and Hazards

The use of blood components that are leukocyte reduced at the bedside may cause unexpected severe hypotension in some recipients, particularly those taking angiotensin-converting enzyme inhibitor medication.

Specific Leukocyte-Reduced Components

All components resulting from the leukocyte reduction process will bear the labeling attribute "leukocytes reduced."

Irradiation

Description

Blood components that contain viable lymphocytes may be irradiated to prevent proliferation of T lymphocytes, which is the immediate cause of TA-GVHD. Irradiated blood is prepared by exposing the component to a radiation source. The standard dose of gamma or x-ray irradiation is 2500 centigray (cGy) targeted to the central portion of the container with a minimum dose of 1500 cGy delivered to any part of the component.

Indications

Irradiated cellular components (whole blood, red cells, platelets, granulocytes, liquid plasma) are indicated for use in patient groups that are at risk for TA-GVHD. At-risk groups include fetal and neonatal recipients of intrauterine transfusions, selected immunocompromised recipients, recipients of cellular components known to be from a blood relative, recipients who have undergone peripheral blood progenitor cell transplantation, recipients of cellular components whose donor is selected for HLA compatibility, and recipients of granulocyte transfusions. Transfused patients receiving purine analogues (eg, fludarabine, cladribine) or certain other biological immunomodulators (eg, alemtuzumab, antithymocyte globulin) may be at risk for TA-GVHD, depending on clinical factors and the source of the biological agent.

Side Effects and Hazards

Irradiation induces erythrocyte membrane damage. Irradiated red cells have been shown to have higher supernatant potassium levels than nonirradiated red cells. Removal of residual supernatant plasma before transfusion may reduce the risks associated with elevated plasma potassium. The expiration date of irradiated red cells is changed to 28 days after irradiation if remaining shelf life exceeds 28 days. There are no known adverse effects following irradiation of platelets and liquid plasma; the expiration date is unchanged.

Specific Irradiated Components

All components that have been irradiated will bear the labeling attribute “irradiated.”

Washing

Description

Washed components are typically prepared using 0.9% Sodium Chloride, Injection USP with or without small amounts of dextrose. Washing removes unwanted plasma proteins, including antibodies and glycerol from previously frozen units. There will also be some loss of red cells and platelets, as well as a loss of platelet function through platelet activation. The shelf life of washed components is no more than 24 hours at 1 to 6 C or 4 hours at 20 to 24 C. Washing is not a substitute for leukocyte reduction, and only cellular components should be washed.

Indications

Washing may be used to reduce exposure to plasma proteins, acellular constituents, or additives (such as mannitol). It is indicated to reduce exposure to antibodies targeting known recipient antigens (such as an Apheresis Platelet unit containing incompatible plasma collected from a mother for the treatment of neonatal alloimmune thrombocytopenia), or to remove constituents that predispose patients to significant or repeated transfusion reactions (eg, removal of IgA-containing plasma in providing transfusion support for an IgA-deficient recipient or in rare recipients experiencing anaphylactoid/anaphylactic reactions to other plasma components).

Specific Washed Components

WASHED RED BLOOD CELLS

WASHED APHERESIS RED BLOOD CELLS

WASHED PLATELETS Ω

WASHED APHERESIS PLATELETS Ω

WASHED APHERESIS PLATELETS PLATELET ADDITIVE SOLUTION ADDED LEUKOCYTES REDUCED Ω

Volume Reduction

Description

Volume reduction is a special manipulation of cellular blood products using centrifugation to remove plasma and storage media in order to concentrate the product. It is typically performed immediately prior to issuing the product for patient administration. The process involves the aseptic removal of a portion of the supernatant, con-

taining plasma and storage medium. Volume reduction removes excess plasma, thereby reducing unwanted plasma proteins, including antibodies. It is more commonly used in pediatric and intrauterine transfusions. There will be some loss of platelet function through platelet activation as a result of volume reduction. The shelf life of volume-reduced components is no more than 24 hours at 1 to 6 C or 4 hours at 20 to 24 C.

Indications

Reducing the plasma volume of cellular components is indicated in cases where consequences of hypervolemia are of concern (such as in infants with compromised cardiac function). Component volume reduction is also used to mitigate adverse transfusion reactions such as TACO, severe allergic reactions, and ABO incompatibilities.

Contraindications

Volume reduction is not a substitute for washing or for dosing with small aliquots.

Specific Volume-Reduced Components

RED BLOOD CELLS PLASMA REDUCED Ω

RED BLOOD CELLS SUPERNATANT REDUCED Ω

APHERESIS RED BLOOD CELLS PLASMA REDUCED Ω

APHERESIS RED BLOOD CELLS SUPERNATANT REDUCED Ω

PLATELETS PLASMA REDUCED Ω

APHERESIS PLATELETS PLASMA REDUCED Ω

APHERESIS PLATELETS PLATELET ADDITIVE SOLUTION ADDED LEUKOCYTES REDUCED SUPERNATANT REDUCED Ω

Additional Testing

This section addresses additional testing performed on previously described blood components. The testing described in this section includes identification of CMV-seronegative blood and identification of low-titer anti-A and/or anti-B blood products.

Identification of CMV-Seronegative Blood

Description

CMV-seronegative blood is selected by testing for antibodies to CMV. Transmission of CMV disease is associated with cellular blood components, specifically those containing mononuclear leukocytes from donors with a history of CMV infection. Plasma, cryoprecipitate, and other plasma-derived blood components are not associated with CMV transmission. Therefore, CMV testing is not necessary for these components.

Indications

In the latently infected donor, CMV is exclusively associated with mononuclear leukocytes. Current studies indicate that transfusion of prestorage leukocyte-reduced blood products safely reduces the risk of CMV transmission to levels not significantly different to transfusion with CMV-seronegative blood. Thus, prestorage leukocyte-reduced components are considered a suitable alternative to CMV-seronegative transfusion.

Transfusion of prestorage leukocyte-reduced or CMV-seronegative blood is indicated in CMV-seronegative recipients who are at risk for severe CMV infections. These at-risk groups include pregnant women and their fetuses, low-birthweight infants, hematopoietic progenitor cell transplant recipients, solid-organ transplant recipients, severely immunosuppressed recipients, and HIV-infected patients.

Identification of Low-Titer Anti-A and/or Anti-B Blood Products

Description

Plasma, apheresis platelets, and whole blood products containing defined titers of anti-A and/or anti-B may reduce the risk of hemolytic transfusion reactions when transfusing ABO-incompatible blood products. Titers considered “low” are not standardized; there is no “safe” titer because hemolytic reactions have been observed at even low titers, with no direct correlation of titer and risk of reactions. Facilities must have policies and procedures to define cutoffs for anti-A and/or anti-B titers for ABO-incompatible blood components.

Table 7. Summary Chart of Blood Components

Category	Major Indications	Action/Recipient Benefit	Not Indicated for	Special Precautions	Hazards*	Rate of Infusion
Red Blood Cells	Symptomatic anemia; red cell exchange transfusion.	Increases oxygen-carrying capacity.	Pharmacologically treatable anemia. Volume expansion.	Must be ABO compatible.	Infectious diseases. Hemolytic, septic/toxic, allergic, febrile reactions. Iron overload. TACO. TRALI. TA-GVHD.	As fast as patient can tolerate but less than 4 hours.
Deglycerolized Red Blood Cells	See Red Blood Cells. IgA deficiency with anaphylactoid/anaphylactic reaction.	See Red Blood Cells. Deglycerolization removes plasma proteins. Risk of allergic and febrile reactions reduced.	See Red Blood Cells.	See Red Blood Cells.	See Red Blood Cells. Hemolysis due to incomplete deglycerolization can occur.	See Red Blood Cells.
Red Blood Cells Leukocytes Reduced	See Red Blood Cells. Reduction of febrile reactions, HLA allo-immunization and CMV infection.	See Red Blood Cells.	See Red Blood Cells. Leukocyte reduction should not be used to prevent TA-GVHD.	See Red Blood Cells.	See Red Blood Cells. Hypotensive reaction may occur if bedside leukocyte reduction filter is used.	See Red Blood Cells.
Washed Red Blood Cells	See Red Blood Cells. IgA deficiency with anaphylactoid/anaphylactic reaction. Recurrent severe allergic reactions to unwashed red cell products.	See Red Blood Cells. Washing reduces plasma proteins. Risk of allergic reactions is reduced.	See Red Blood Cells. Washing is not a substitute for leukocyte reduction.	See Red Blood Cells.	See Red Blood Cells.	See Red Blood Cells.

(Continued)

Table 7. Summary Chart of Blood Components (Continued)

Category	Major Indications	Action/Recipient Benefit	Not Indicated for	Special Precautions	Hazards*	Rate of Infusion
Whole Blood	Symptomatic anemia with large volume deficit. Treat the massively bleeding patient.	Increases oxygen-carrying capacity. Increases blood volume. Variably contributes plasma coagulation factors and platelets.	Condition responsive to specific component. Treatment of coagulopathy.	Must be ABO identical or as defined by local policies and procedures.	See Red Blood Cells.	As fast as patient can tolerate but less than 4 hours.
Fresh Frozen Plasma (FFP)	Clinically significant plasma protein deficiencies when no specific coagulation factor concentrates are available. TTP.	Source of plasma proteins, including all coagulation factors.	Volume expansion. Coagulopathy that can be more effectively treated with specific therapy. Correcting a minimally elevated INR.	Must be ABO compatible.	Infectious diseases. Allergic, febrile reactions. TACO. TRALI.	Less than 4 hours.
Plasma Frozen Within 24 Hours After Phlebotomy (PF24)	Clinically significant deficiency of stable coagulation factors when no specific coagulation factor concentrates are available. TTP.	Source of nonlabile plasma proteins. Levels of Factor VIII are significantly reduced and levels of Factor V and other labile plasma proteins are variable compared to FFP.	Volume expansion. Deficiencies of labile coagulation factors, including Factors V and VIII and Protein C. See FFP.	Must be ABO compatible.	See FFP.	Less than 4 hours.

Plasma Frozen Within 24 Hours After Phlebotomy Held At Room Temperature Up To 24 Hours After Phlebotomy (PF24RT24)	Clinically significant deficiency of stable coagulation factors when no specific coagulation factor concentrates are available. TTP.	Source of nonlabile plasma proteins. Levels of Factor V, Factor VIII, and Protein S are significantly reduced, and levels of other labile plasma proteins are variable compared with FFP.	Volume expansion. Deficiencies of labile coagulation factors, including Factors V and VIII and Protein S. See Fresh Frozen Plasma (FFP).	Must be ABO compatible.	See FFP.	Less than 4 hours.
Plasma Cryoprecipitate Reduced	TTP.	Plasma protein replacement for plasma exchange in TTP. Deficient in fibrinogen, vWF, Factors VIII and XIII. Deficient in high-molecular-weight vWF multimers as compared to FFP.	Volume expansion. Deficiency of coagulation factors known to be depleted in this product: fibrinogen, vWF, Factors VIII and XIII.	Must be ABO compatible.	See FFP.	Less than 4 hours.
Thawed Plasma Ω	Clinically significant deficiency of stable coagulation factors when no specific coagulation factor concentrates are available. TTP.	Source of plasmaproteins. Levels and activation state of coagulation proteins in thawed plasma are variable and change over time.	Not indicated as treatment for isolated coagulation factor deficiencies or specific plasma protein deficiencies.	Must be ABO compatible.	See FFP.	Less than 4 hours.

(Continued)

Table 7. Summary Chart of Blood Components (Continued)

Category	Major Indications	Action/Recipient Benefit	Not Indicated for	Special Precautions	Hazards*	Rate of Infusion
Thawed Plasma Cryoprecipitate Reduced Ω	TTP.	Plasma protein replacement for plasma exchange in TTP. Deficient in fibrinogen, vWF, Factors VIII and XIII.	Volume expansion. Deficiency of coagulation factors known to be depleted in this product: fibrinogen, vWF, Factors VIII and XIII. See FFP.	Must be ABO compatible.	See FFP.	Less than 4 hours.
Liquid Plasma	Initial treatment of patients undergoing massive transfusion.	Coagulation support for life-threatening trauma/hemorrhages. The profile of plasma proteins in Liquid Plasma is not completely characterized. Levels and activation state of coagulation proteins are dependent on production methods and storage.	Not indicated as treatment for coagulation factor deficiencies where other products are available with higher factor concentrations. See FFP.	Must be ABO compatible.	See FFP.	Less than 4 hours.

Cryoprecipitated AHF; Pooled Cryoprecipitated AHF	Hypofibrinogenemia. Factor XIII deficiency. Second-line therapy of von Willebrand disease, hemophilia A, and uremic bleeding.	Provides fibrinogen, vWF, Factors VIII and XIII.	Not indicated if specific concentrates are available. Deficiency of any plasma protein other than those enriched in Cryoprecipitated AHF.		Infectious diseases. Allergic, febrile reactions.	Less than 4 hours.
Platelets/Apheresis Platelets	Bleeding due to thrombocytopenia or platelet function abnormality, including antiplatelet drugs. Prevention of bleeding from marrow hypoplasia.	Improves hemostasis. Apheresis platelets may be HLA (or other antigen) selected.	Plasma coagulation deficits. Some conditions with rapid platelet destruction (eg, ITP, TTP) unless life-threatening hemorrhage.	Should use only platelet-compatible filters (check manufacturer's instructions).	Infectious diseases. Septic/toxic, allergic, febrile reactions. TACO. TRALI. TA-GVHD.	Less than 4 hours.
Platelets Leukocytes Reduced/ Apheresis Platelets Leukocytes Reduced	See Platelets. Reduction of febrile reactions, HLA alloimmunization and CMV infection.	See Platelets.	See Platelets. Leukocyte reduction should not be used to prevent TA-GVHD.	See Platelets.	See Platelets.	See Platelets.
Apheresis Platelets Platelet Additive Solution Added Leukocytes Reduced	See Platelets Leukocytes Reduced.	See Platelets.	See Platelets Leukocytes Reduced.	See Platelets.	See Platelets.	See Platelets.

(Continued)

Table 7. Summary Chart of Blood Components (Continued)

Category	Major Indications	Action/Recipient Benefit	Not Indicated for	Special Precautions	Hazards*	Rate of Infusion
Apheresis Granulocytes Ω	Neutropenia with infection, unresponsive to appropriate antibiotics.	Provides granulocytes and platelets.	Infection responsive to antibiotics; eventual marrow recovery not expected.	Must be ABO compatible. Use only filters specifically approved by a manufacturer for granulocyte transfusions (check manufacturer's instructions).	Infectious diseases. Hemolytic, allergic, febrile reactions. TACO. TRALI. TA-GVHD. Maintain caution. Pulmonary reactions may occur in patients receiving concomitant amphotericin B.	One unit over 2-4 hours. Closely observe for reactions.

*For all cellular components there is a risk the recipient may become alloimmunized and experience rapid destruction of certain types of blood products. Red-cell-containing components and thawed plasma (thawed FFP, thawed PF24, thawed PF24RT24, or Thawed Plasma) should be stored at 1 to 6 C. Platelets, Granulocytes, and thawed Cryoprecipitate should be stored at 20 to 24 C. Disclaimer: Please check the corresponding section of the *Circular* for more detailed information.

AHF = antihemophilic factor; CMV = cytomegalovirus; HLA = human leukocyte antigen; ITP = immune thrombocytopenic purpura; IUT = intrauterine transfusion; TA-GVHD = transfusion-associated graft-vs-host disease; TACO = transfusion-associated circulatory overload; TRALI = transfusion-related acute lung injury; TTP = thrombotic thrombocytopenic purpura; vWF = von Willebrand Factor.

Refer to the *United States Industry Consensus Standard for the Uniform Labeling of Blood and Blood Components Using ISBT 128* (<https://www.fda.gov/regulatory-information/search-fda-guidance-documents/united-states-industry-consensus-standard-uniform-labeling-blood-and-blood-components-using-isbt-128>) for labeling recommendations. Options include placing the titer value on a tie tag.

Indications

Group O Whole Blood and group A plasma tested for anti-A and/or anti-B may be used as an initial resuscitation fluid for an acutely bleeding patient prior to determining the recipient blood group.

The transfusing facility must have policies and procedures in place addressing specific indications for use, product specifications, administration instructions, and a defined maximum number of units to be transfused to each patient.

Contraindications

ABO-incompatible products should not be transfused when an appropriate product that is ABO compatible is readily available, or when the risk of administering ABO-incompatible blood components outweighs the potential therapeutic benefit.

References

AABB References

- Cohn CS, Delaney M, Johnson ST, et al, eds. Technical manual. 21st ed. Bethesda, MD: AABB, 2023.
- Galel SA, ed. Standards for blood banks and transfusion services. 34th ed. Bethesda, MD: AABB, 2024.
- Kopko PM, ed. Transfusion reactions. 5th ed. Bethesda, MD: AABB Press, 2021.
- Mo Y, Roseff SD, Wong ECC, eds. Pediatric hemotherapy data card. 5th ed. Bethesda, MD: AABB, 2020.
- Wong ECC, Roseff SD, Bandarenko N, eds. Pediatric transfusion: A handbook. 5th ed. Bethesda, MD: AABB, 2020.

FDA References

- FDA: Code of federal regulations. Title 21. Washington DC: US Government Publishing Office, 2023. [Available at <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/cfrsearch.cfm> (accessed October 4, 2023).]
- FDA: eCFR. Title 21. Washington DC: US Government Publishing Office, updated September 26, 2023. [Available at <https://www.ecfr.gov/current/title-21> (accessed October 4, 2023).]
- FDA: Blood guidances. [Available at <https://www.fda.gov/vaccines-blood-biologics/biologics-guidances/blood-guidances> (accessed October 4, 2023).]

General

- American Society of Anesthesiologists Task Force on Perioperative Blood Transfusion and Adjuvant Therapies. Practice guidelines for perioperative blood transfusion and adjuvant therapies: An updated report. *Anesthesiology* 2015;122:241-75.
- Chou ST, Alsawas M, Fasano RM, et al. American Society of Hematology 2020 guidelines for sickle cell disease: Transfusion support. *Blood Adv* 2020;4(2):327-55.
- Desmet L, Lacroix J. Transfusion in pediatrics. *Crit Care Clin* 2004;20:299-311.
- Expert Working Group. Guidelines for red blood cell and plasma transfusion for adults and children. *CMAJ* 1997;156(11 Suppl):S1-S24.
- Ferraris VA, Brown JB, Despotis GJ, et al. 2011 Update to the Society of Thoracic Surgeons and the Society of Cardiovascular Anesthesiologists blood conservation clinical practice guidelines. *Ann Thorac Surg* 2011;91:944-82.
- Gajewski JL, Johnson VV, Sandler SG, et al. A review of transfusion practice before, during and after hematopoietic progenitor cell transplantation. *Blood* 2008;112:3036-47.
- Harvey AR, Basavaraju SV, Chung KW, Kuehnert MJ. Transfusion-related adverse reactions reported to the National Healthcare Safety Network Hemovigilance Module, United States, 2010 to 2012. *Transfusion* 2015;55:709-18.
- Kleinman S, Chan P, Robillard P. Risks associated with transfusion of cellular blood components in Canada. *Transfus Med Rev* 2003;17:120-62.
- McFarland JG. Perioperative blood transfusions. *Chest* 1999;115:113S-21S.

- Murdock J, Watson D, Doree CJ, et al. Drugs and blood transfusions: Dogma or evidence-based practice? *Transfus Med* 2009;19:6-15.
- New HV, Berryman J, Bolton-Maggs PH, et al. British Committee for Standards in Haematology. Guidelines on transfusion for fetuses, neonates and older children. *Br J Haematol* 2016;175:784-828.
- New HV, Stanworth SJ, Gottstein R, et al on behalf of the BSH Guidelines Transfusion Task Force. British Society for Haematology Guidelines on transfusion for fetuses, neonates and older children. *Br J Haematol* 2016;175:784-828. Addendum 2020;191:725-7.
- Roseff SD, Luban NL, Manno CS. Guidelines for assessing appropriateness of pediatric transfusion. *Transfusion* 2002;42:1398-413.
- Sazama K, DeChristopher PJ, Dodd R, et al. Practice parameter for the recognition, management, and prevention of adverse consequences of blood transfusion. *Arch Pathol Lab Med* 2000;124:61-70.
- Valentine SL, Bembea MM, Muszynski JA, et al. Consensus recommendations for red blood cell transfusion practice in critically ill children from the Pediatric Critical Care Transfusion and Anemia Expertise Initiative. *Pediatr Crit Care Med* 2018;19(9):884-98.
- Wortham ST, Ortolano GA, Wenz B. A brief history of blood filtration: Clot screens, microaggregate removal and leukocyte reduction. *Transfus Med Rev* 2003;17:216-22.

Infectious Complications

- AABB, Clinical Transfusion Medicine Committee; Heddle NM, Boeckh M, et al. AABB committee report: Reducing transfusion-transmitted cytomegalovirus infections. *Transfusion* 2016;56:1581-7.
- Alter HJ, Stramer SL, Dodd RY. Emerging infectious diseases that threaten the blood supply. *Semin Hematol* 2007;44:32-41.
- Benjamin RJ, Kline L, Dy BA, et al. Bacterial contamination of whole-blood-derived platelets: The introduction of sample diversion and prestorage pooling with culture testing in the American Red Cross. *Transfusion* 2008;48:2348-55.
- Bloch EM, Krause PJ, Tonnetti L. Preventing transfusion-transmitted babesiosis. *Pathogens* 2021;10(9):1176.
- Crowder LA, Schonberger LB, Dodd RY, et al. Creutzfeldt-Jakob disease lookback study: 21 Years of surveillance for transfusion transmission risk. *Transfusion* 2017;57:1875-8.
- Dodd RY, Foster GA, Stramer SL. Keeping blood transfusion safe from West Nile virus: American Red Cross experience, 2003 to 2012. *Transfus Med Rev* 2015;29(3):153-61.
- Domanović D, Gossner CM, Lieshout-Krikke R, et al. West Nile and Usutu virus infections and challenges to blood safety in the European Union. *Emerg Infect Dis* 2019;25(6):1050-7.
- Dorsey K, Zou S, Schonberger LB, et al. Lack of evidence of transfusion transmission of Creutzfeldt-Jakob disease in a US surveillance study. *Transfusion* 2009;49:977-84.
- Eder AF, Chambers LA. Noninfectious complications of blood transfusion. *Arch Pathol Lab Med* 2007;131:708-18.
- Eder AF, Kennedy JM, Dy BA, et al. Limiting and detecting bacterial contamination of apheresis platelets: Inlet-line diversion and increased culture volume improve component safety. *Transfusion* 2009;49:1154-63.
- Fang D, McCullough J. Transfusion-transmitted *Babesia microti*. *Transfus Med Rev* 2016;30(3):132-8.
- Katz LM, Spencer BR. Travel and related health history: Malaria, prions, and other transfusion-transmitted diseases. In: Eder A, Goldman M, eds. *Screening blood donors with the donor history questionnaire*. Bethesda, MD: AABB Press, 2019.
- Kracalik I, Kent AG, Villa CH, et al. Posttransfusion sepsis attributable to bacterial contamination in platelet collection set manufacturing facility, United States. *Emerg Infect Dis* 2023;29(10):1979-89.
- Moritz ED, Winton CS, Tonnetti L, et al. Screening for *Babesia microti* in the U.S. blood supply. *N Engl J Med* 2016;375:2236-45.
- Mungai M, Tegtmeier G, Chamberland M, Parise M. Transfusion-transmitted malaria in the United States from 1963 through 1999. *N Engl J Med* 2001;344(26):1973-8.
- O'Brien SF, Delage G, Seed CR, et al. The epidemiology of imported malaria and transfusion policy in 5 nonendemic countries. *Transfus Med Rev* 2015;29(3):162-71.
- Petersen LR, Busch MP. Transfusion-transmitted arboviruses. *Vox Sang* 2010;98:495-503.
- Pozzo di Borgo A, Rochette S, Gausson A, et al. Transmission of variant Creutzfeldt-Jakob disease through blood transfusion and plasma-derived products: A narrative review of observed and modeled risks. *Transfus Med Rev* 2023;37(3):150747.

- Seed CR, Hewitt PE, Dodd RY, et al. Creutzfeldt-Jakob disease and blood transfusion safety. *Vox Sang* 2018;113(3):220-31.
- Stramer SL. Reacting to an emerging safety threat: West Nile virus in North America. *Dev Biol (Basel)* 2007;127:43-58.
- Stramer SL, Glynn SA, Kleinman SH, et al. Detection of HIV-1 and HCV infections among antibody-negative blood donors by nucleic-acid amplification testing. *N Engl J Med* 2004;351:760-8.
- Stramer SL, Hollinger B, Katz LM, et al. Emerging infectious disease agents and their potential threat to transfusion safety. *Transfusion* 2009;49(Suppl 2):1S-29S.
- Vamvakas EC. Is white blood cell reduction equivalent to antibody screening in preventing transmission of cytomegalovirus by transfusion? A review of the literature and meta-analysis. *Transfus Med Rev* 2005;19:181-99.
- Zou S, Stramer SL, Dodd RY. Donor testing and risk: Current prevalence, incidence and residual risk of transfusion-transmissible agents in US allogeneic donations. *Transfus Med Rev* 2012;26:119-28.

Pathogen Reduction Technology

- Cid J. Prevention of transfusion-associated graft-versus-host disease with pathogen-reduced platelets with amotosalen and ultraviolet A light: A review. *Vox Sang* 2017;112:607-13.
- Cid J, Caballo C, Pino M, et al. Quantitative and qualitative analysis of coagulation factors in cryoprecipitate prepared from fresh-frozen plasma inactivated with amotosalen and ultraviolet A light. *Transfusion* 2013;53:600-5.
- Cushing M, Asmis L, Harris R, et al. Efficacy of a new pathogen-reduced cryoprecipitate stored 5 days after thawing to correct dilution coagulopathy in vitro. *Transfusion* 2019;59:1818-26.
- Thomas KA, Shea SM, Spinella PC. Effects of pathogen reduction technology and storage duration on the ability of cryoprecipitate to rescue induced coagulopathies in vitro. *Transfusion* 2021;61(6):1943-54.

Malaria

- Kitchen AD, Chiodini PL. Malaria and blood transfusion. *Vox Sang* 2006;90:77-84.
- Mungai M, Tegtmeier G, Chamberland M, et al. Transfusion-transmitted malaria in the United States from 1963 through 1999. *N Engl J Med* 2001;344:1973-8.
- Verra F, Angheben A, Martello E, et al. A systematic review of transfusion-transmitted malaria in non-endemic areas. *Malaria J* 2018;17:36.

Transfusion-Associated Graft-vs-Host Disease (TA-GVHD)/Irradiation

- Bahar B, Tormey CA. Prevention of transfusion-associated graft-versus-host disease with blood product irradiation: The past, present and future. *Arch Pathol Lab Med* 2018;142:662-7.
- Dwyre DM, Holland PV. Transfusion-associated graft-versus-host disease. *Vox Sang* 2008;95:85-93.
- Foukaneli T, Kerr P, Bolton-Maggs PHB, et al. Guidelines on the use of irradiated blood components. *Br J Haematol* 2020;191(5):704-24.
- Kopolovic I, Ostro J, Tsubota H, et al. A systematic review of transfusion-associated graft-versus-host disease. *Blood* 2015;126:406-14.
- Przepiorka D, LeParc GF, Stovall MA, et al. Use of irradiated blood components. Practice parameter. *Am J Clin Pathol* 1996;106:6-11.
- Ruhl H, Bein G, Sachs UJH. Transfusion-associated graft-versus-host disease. *Transfus Med Rev* 2009;23:62-71.

Transfusion-Related Acute Lung Injury (TRALI)

- Eder AF, Herron RM Jr, Strupp A, et al. Effective reduction of transfusion-related acute lung injury risk with male-predominant plasma strategy in the American Red Cross (2006-2008). *Transfusion* 2010;50:1732-42.
- Goldman M, Webert KE, Arnold DM, et al. Proceedings of a consensus conference: Towards an understanding of TRALI. *Transfus Med Rev* 2005;19:2-31.
- Otrock ZK, Liu C, Grossman BJ. Transfusion-related acute lung injury risk mitigation: An update. *Vox Sang* 2017;112:694-703.
- Rana R, Fernandez-Perez ER, Khan SA, et al. Transfusion-related acute lung injury and pulmonary edema in critically ill patients: A retrospective study. *Transfusion* 2006;46:1478-83.
- Sanchez R, Toy P. Transfusion-related acute lung injury: A pediatric perspective. *Pediatr Blood Cancer* 2005;45:248-55.
- Vlaar APJ, Toy P, Fung M, et al. A consensus redefinition of transfusion-related acute lung injury. *Transfusion* 2019;59:2465-76.

Transfusion-Associated Circulatory Overload (TACO)

- Andrzejewski C Jr, Casey MA, Popovsky MA. How we view and approach transfusion-associated circulatory overload: Pathogenesis, diagnosis, management, mitigation, and prevention. *Transfusion* 2013;53:3037-47.
- Bosboom JJ, Klanderman RB, Migdady Y, et al. Transfusion-associated circulatory overload: A clinical perspective. *Trans Med Rev* 2019;33:69-77.
- Gajic O, Gropper MA, Hubmayr RD. Pulmonary edema after transfusion: How to differentiate transfusion-associated circulatory overload from transfusion-related acute lung injury. *Crit Care Med* 2006;34(5 Suppl):S109-13.
- Klanderman RB, Bosboom JJ, Migdady Y, et al. Transfusion-associated circulatory overload—a systematic review of diagnostic biomarkers. *Transfusion* 2019;59:795-805.
- Popovsky MA. Transfusion and the lung: Circulatory overload and acute lung injury. *Vox Sang* 2004;87(Suppl 2):62-5.
- Roubinian NH, Hendrickson JE, Triulzi DJ, et al. Incidence and clinical characteristics of transfusion-associated circulatory overload using an active surveillance algorithm. *Vox Sang* 2017;112:56-63.
- Semple JW, Rebetz J, Kapur R. Transfusion-associated circulatory overload and transfusion-related acute lung injury. *Blood* 2019;133:1840-53.
- Wiersum-Osselton JC, Whitaker B, Grey S, et al. Revised international surveillance case definition of transfusion-associated circulatory overload: A classification agreement validation study. *Lancet Haematol* 2019;6:e350-8.

Febrile Nonhemolytic Transfusion Reactions

- Geiger TL, Howard SC. Acetaminophen and diphenhydramine premedication for allergic and febrile nonhemolytic transfusion reactions: Good prophylaxis or bad practice? *Transfus Med Rev* 2007;21:1-12.
- King KE, Shirey RS, Thoman SK, et al. Universal leukoreduction decreases the incidence of febrile non-hemolytic transfusion reactions to RBCs. *Transfusion* 2004;44:25-9.
- Menis M, Forshee RA, Anderson SA, et al. Febrile non-haemolytic transfusion reaction occurrence and potential risk factors among the U.S. elderly transfused in the inpatient setting, as recorded in Medicare databases during 2011-2012. *Vox Sang* 2015;108(3):251-61.
- Paglino JC, Pomper GJ, Fisch GS, et al. Reduction of febrile but not allergic reactions to RBCs and platelets after conversion to universal prestorage leukoreduction. *Transfusion* 2004;44:16-24.
- Wang H, Ren D, Sun H, Liu J. Research progress on febrile non-hemolytic transfusion reaction: A narrative review. *Ann Transl Med* 2022;10(24):1401.

Platelet Refractoriness

- Blanchette VS, Johnson J, Rand M. The management of alloimmune neonatal thrombocytopenia. *Baillieres Best Pract Res Clin Haematol* 2000;13:365-90.
- Dzik S. How I do it: Platelet support for refractory patients. *Transfusion* 2007;47:374-8.
- Hod E, Schwartz J. Platelet transfusion refractoriness. *Br J Haematol* 2008;142:348-60.
- Juskewitch JE, Norgan AP, De Goey SR, et al. How do I...manage the platelet transfusion-refractory patient? *Transfusion* 2017;57:2828-35.
- Kiefel V, Bassler D, Kroll H, et al. Antigen-positive platelet transfusion in neonatal alloimmune thrombocytopenia (NAIT). *Blood* 2006;107:3761-3.
- McVey M, Cserti-Gazdewich CM. Platelet transfusion refractoriness responding preferentially to single donor aphaeresis platelets compatible for both ABO and HLA. *Transfus Med* 2010;20:346-53.
- Refaai M, Phipps R, Spinelli S, et al. Platelet transfusions: Impact on hemostasis, thrombosis, inflammation and clinical outcomes. *Thromb Res* 2011;10:1012-16.
- Sacher RA, Kickler TS, Schiffer CA, et al for the CAP Transfusion Medicine Resource Committee. Management of patients refractory to platelet transfusion. *Arch Pathol Lab Med* 2003;127:409-14.
- Schmidt AE, Refaai MA, Coppage M. HLA-mediated platelet refractoriness: An ACLPS critical review. *Am J Clin Pathol* 2019;151:353-63.
- Slichter SJ, Davis K, Enright H, et al. Factors affecting post transfusion platelet increments, platelet refractoriness and platelet transfusion intervals in thrombocytopenic patients. *Blood* 2006;105:4106-14.

Citrate Toxicity

- Dzik WH, Kirkley SA. Citrate toxicity during massive blood transfusion. *Transfus Med Rev* 1988;2:76-94.
- Monchi M. Citrate pathophysiology and metabolism. *Transfus Apher Sci* 2017;56:28-30.
- Sihler KC, Napolitano LM. Complications of massive transfusion. *Chest* 2010;137:209-20.

Allergic and Anaphylactoid/Anaphylactic Reactions

- Marti-Carvajal AJ, Sola I, Gonzalez LE, et al. Pharmacological interventions for the prevention of allergic and febrile non-haemolytic transfusion reactions. *Cochrane Database Syst Rev* 2010;6:CD007539.
- Sandler SG. How I manage patients suspected of having had an IgA anaphylactic transfusion reaction. *Transfusion* 2006;46:10-13.
- Sandler SG, Eder A, Goldman M, et al. The entity of immunoglobulin A-related anaphylactic transfusion reactions is not evidence based. *Transfusion* 2015;55:199-204.
- Savage W, Tobian A, Savage J, et al. Scratching the surface of allergic transfusion reactions. *Transfusion* 2013;53: 1361-71.
- Savage W, Tobian A, Savage J, et al. Transfusion and component characteristics are not associated with allergic transfusion reactions to apheresis platelets. *Transfusion* 2015;55:296-300.
- Tobian AAR, Savage WJ, Tisch DJ, et al. Prevention of allergic transfusion reactions to platelets and red blood cells through plasma reduction. *Transfusion* 2011;51:1676-83.
- Vassallo RR. Review: IgA anaphylactic transfusion reactions. Part I. Laboratory diagnosis, incidence, and supply of IgA-deficient products. *Immunohematol* 2004;20:226-33.

Iron Overload

- Coates TD, Wood JC. How we manage iron overload in sickle cell patients. *Br J Haematol* 2017;117:703-16.
- Majhail NS, Lazarus HM, Burns LJ. Iron overload in hematopoietic cell transplantation. *Bone Marrow Transplant* 2008;41:997-1003.
- Shander A, Sazama K. Clinical consequences of iron overload from chronic red blood cell transfusions, its diagnosis, and its management by chelation therapy. *Transfusion* 2010;50:1144-55.
- Taher AT, Saliba AN. Iron overload in thalassemia: Different organs at different rates. *Hematology Am Soc Hematol Educ Program* 2017;2017(1):265-71.

Whole Blood

- Leeper CM, Yazer MH, Cladis FP, et al. Use of uncrossmatched cold-stored whole blood in injured children with hemorrhagic shock. *JAMA Pediatr* 2018;172:491-2.
- Leeper CM, Yazer MH, Triulzi DJ, et al. Whole blood is superior to component transfusion for injured children: A propensity matched analysis. *Ann Surg* 2020;272:590-4.
- Morgan KM, Yazer MH, Triulzi DJ, et al. Safety profile of low-titer group O whole blood in pediatric patients with massive hemorrhage. *Transfusion* 2021;61(Suppl 1):S8-S14.
- Sivertsen J, Braathen H, Lunde THF, et al. Preparation of leuko-reduced whole blood for transfusion in austere environments; effects of forced filtration, storage agitation, and high temperatures on hemostatic function. *J Trauma Acute Care Surg* 2018;84(6S Suppl 1):S93-S103.
- Spinella PC, Perkins JG, Trathwohl KW, et al. Warm fresh whole blood is independently associated with improved survival for patients with combat-related traumatic injuries. *J Trauma* 2009;66(4 Suppl):S69-76.
- Williams J, Merutka N, Meyer D, et al. Safety profile and impact of low-titer group O whole blood for emergency use in trauma. *J Trauma Acute Care Surg* 2020;88:87-93.

Red Blood Cells

- Bell EF, Strauss RG, Widness JA, et al. Randomized trial of liberal versus restrictive guidelines for red blood cell transfusion in preterm infants. *Pediatrics* 2005;115:1685-91.
- Cable CA, Razavi SA, Roback JD, Murphy DJ. RBC transfusion strategies in the ICU: A concise review. *Crit Care Med* 2019;47(11):1637-44.
- Carless PA, Henry DA, Carson JL, et al. Transfusion thresholds and other strategies for guiding allogeneic red blood cell transfusion (review). *Cochrane Database Syst Rev* 2010;10:CD002042.
- Carson JL, Brooks MM, Abbott JD, et al. Liberal versus restrictive transfusion thresholds for patients with symptomatic coronary artery disease. *Am Heart J* 2013;165:964-71.e1.
- Carson JL, Grossman BJ, Kleinman S, et al for the AABB Clinical Transfusion Medicine Committee. Red blood cell transfusion: A clinical practice guideline from the AABB. *Ann Intern Med* 2012;157:49-58.
- Carson JL, Guyatt G, Heddle NM, et al. Clinical practice guidelines from the AABB: Red blood cell transfusion thresholds and storage. *JAMA* 2016;316:2025-35.
- Carson JL, Stanworth SJ, Guyatt G, et al. Red blood cell transfusion: 2023 AABB international guidelines. *JAMA* 2023;330(19):1892-902.
- Carson JL, Terrin ML, Novek H, et al. Liberal or restrictive transfusion in high-risk patients after hip surgery. *N Engl J Med* 2011;365:2453-62.

- Cooper HA, Rao SV, Greenberg MD, et al. Conservative versus liberal red cell transfusion in acute myocardial infarction (the CRIT Randomized Pilot Study). *Am J Cardiol* 2011;108:1108-11.
- D'Alessandro A, Nemkov T, Hansen K, et al. Red blood cell storage in additive solution-7 preserves energy and redox metabolism: A metabolomics approach. *Transfusion* 2015;55:2955-66.
- Dumont L, Cancelas J, Maes L, et al. Overnight, room temperature hold of whole blood followed by 42-day storage of red blood cells in additive solution-7. *Transfusion* 2015;55:485-90.
- Franz AR, Engel C, Bassler D, et al. Effects of liberal vs restrictive transfusion thresholds on survival and neuro-cognitive outcomes in extremely low-birthweight infants: The ETTNO randomized clinical trial. *JAMA* 2020; 324:560-70.
- Gould S, Cimino MJ, Gerber DR. Packed red blood cell transfusion in the intensive care unit: Limitations and consequences. *Am J Crit Care* 2007;16:39-48.
- Hajjar LA, Vincent JL, Galas FR, et al. Transfusion requirements after cardiac surgery: The TRACS randomized controlled trial. *JAMA* 2010;304:1559-67.
- Hébert PC, Wells G, Blajchman MA, et al. A multicenter, randomized, controlled clinical trial of transfusion requirements in critical care. *N Engl J Med* 1999;340:409-17.
- Hébert PC, Yetisir E, Martin C. Is a low transfusion threshold safe in critically ill patients with cardiovascular disease? *Crit Care Med* 2001;29:227-34.
- Holst LB, Haase N, Wettersley J. Lower versus higher hemoglobin thresholds for transfusion in septic shock. *N Engl J Med* 2014;27:1381-91.
- Istaphanous GK, Wheeler DS, Lisco SJ, et al. Red blood cell transfusion in critically ill children: A narrative review. *Pediatr Crit Care Med* 2011;12:174-83.
- Josephson CD, Su LL, Hillyer KL, et al. Transfusion in the patient with sickle cell disease: A critical review of the literature and transfusion guidelines. *Transfus Med Rev* 2007;21:118-33.
- Kirpalani H, Whyte RK, Andersen C, et al. The Premature Infants in Need of Transfusion (PINT) Study: A randomized, controlled trial of a restrictive (low) versus liberal (high) transfusion threshold for extremely low birth weight infants. *J Pediatr* 2006;149:301-7.
- Lacroix J, Hébert PC, Hutchison JS, et al; TRIPICU Investigators; Canadian Critical Care Trials Group; Pediatric Acute Lung Injury and Sepsis Investigators Network. Transfusion strategies for patients in pediatric intensive care units. *N Engl J Med* 2007;356:1609-19.
- Lavoie J. Blood transfusion risks and alternative strategies in pediatric patients. *Paediatr Anaesth* 2011;21:14-24.
- Lu W, Ziman A, Yan MTS, et al. Serologic reactivity of unidentified specificity in antenatal testing and hemolytic disease of the fetus and newborn: The BEST collaborative study. *Transfusion* 2023;63(4):817-25.
- Luban NL. Management of anemia in the newborn. *Early Hum Dev* 2008;84:493-8.
- Mazer CD, Whitlock RP, Fergusson DA, et al. Restrictive or liberal red-cell transfusion for cardiac surgery. *N Engl J Med* 2017;377:2133-44.
- Mazer CD, Whitlock RP, Fergusson DA, et al. Six-month outcomes after restrictive or liberal transfusion for cardiac surgery. *N Engl J Med* 2018;379:1224-33.
- Murphy GJ, Pike K, Rogers CA, et al. Liberal or restrictive transfusion after cardiac surgery. *N Engl J Med* 2015;372:997-1008.
- Poole J, Daniels G. Blood group antibodies and their significance in transfusion medicine. *Transfus Med Rev* 2007;21:58-71.
- Shah A, Brunskill SJ, Desborough M, et al. Transfusion of red blood cells stored for shorter versus longer duration for all conditions. *Cochrane Database Syst Rev* 2018;12:CD010801.
- Shander A, Fink A, Javidroozi M, et al. Appropriateness of allogeneic red blood cell transfusion: The international consensus conference on transfusion outcomes. *Transfus Med Rev* 2011;25:232-46.
- Stainsby D, MacLennan S, Thomas D, et al for the Standards in Haematology Writing Group. Guidelines for the management of massive blood loss. *Br J Haematol* 2006;135:634-41.
- Veale M, Healey G, Sran A, et al. AS-7 improved in vitro quality of red blood cells prepared from whole blood held overnight at room temperature. *Transfusion* 2015;55:108-14.
- Villanueva C, Colomo A, Bosch A, et al. Transfusion strategies for acute upper gastrointestinal bleeding. *N Engl J Med* 2013;368:11-21.
- Weiskopf RB, Viele MK, Feiner J, et al. Human cardiovascular and metabolic response to acute, severe isovolemic anemia. *JAMA* 1998;279:217-21.

Plasma, Cryoprecipitate, and Granulocytes

- Ahmed S, Harrity C, Johnson S, et al. The efficacy of fibrinogen concentrate compared with cryoprecipitate in major obstetric haemorrhage - an observational study. *Transfus Med* 2012;22:344-9.
- Backholer L, Green L, Huish S, et al. A paired comparison of thawed and liquid plasma. *Transfusion* 2017;57:881-9.
- Blackall DP, Uhl L, Spitalnik SL, et al for the Transfusion Practices Committee. Cryoprecipitate-reduced plasma: Rationale for use and efficacy in the treatment of thrombotic thrombocytopenic purpura. *Transfusion* 2001;41:840-4.
- Bostrom F, Sjö Dahl M, Wehlin L, et al. Coagulation parameters in apheresis and leukodepleted whole-blood plasma during storage. *Transfusion* 2007;47:460-3.
- Burtelov M, Riley E, Druzin M, et al. How we treat: Management of life-threatening primary postpartum hemorrhage with a standardized massive transfusion protocol. *Transfusion* 2007;47:1564-72.
- Callum J, Farkouh ME, Scales DC, et al. Effect of fibrinogen concentrate vs cryoprecipitate on blood component transfusion after cardiac surgery: The FIBRES randomized clinical trial. *JAMA* 2019;322(20):1966-76.
- Callum JL, Karkouti K, Lin Y. Cryoprecipitate: The current state of knowledge. *Transfus Med Rev* 2009;23:177-88.
- Cardigan R, Green L. Thawed and liquid plasma – what do we know? *Vox Sang*, 2015;109:1-10.
- Cardigan R, Lawrie A, Mackie IJ, et al. The quality of fresh-frozen plasma produced from whole blood stored at 4 degrees C overnight. *Transfusion* 2005;45:1342-8.
- Castaman G, Linari SJ. Diagnosis and treatment of von Willebrand disease and rare bleeding disorders. *Clin Med* 2017;6(4):45.
- Caudill JS, Nichols WL, Plumhoff EA, et al. Comparison of coagulation factor XIII content and concentration in cryoprecipitate and fresh-frozen plasma. *Transfusion* 2009;49:765-70.
- Curry N, Rourke C, Davenport R, et al. Early cryoprecipitate for major haemorrhage in trauma: A randomised controlled feasibility trial. *Br J Anaesth* 2015;115:76-83.
- Dara SI, Rana R, Afessa B, et al. Fresh frozen plasma transfusion in critically ill medical patients with coagulopathy. *Crit Care Med* 2005;33:2667-71.
- Droubatchevskaia N, Wong MP, Chipperfield KM, et al. Guidelines for cryoprecipitate transfusion. *BCM J* 2007;49:441-5.
- Dumont L, Cancelas J, Maes L, et al. The bioequivalence of frozen plasma prepared from whole blood held overnight at room temperature compared to fresh-frozen plasma prepared within eight hours of collection. *Transfusion* 2015;55:476-84.
- Elebute M, Massey E, Benjamin S, et al for the NHS Granulocyte Working Group. Clinical Guidelines for the Use of Granulocyte Transfusion. Bristol, UK: National Health Service Blood and Transplant, 2021.
- Gajic O, Dzik WH, Toy P. Fresh frozen plasma and platelet transfusion for nonbleeding patients in the intensive care unit. *Crit Care Med* 2006;34(5 Suppl):S170-3.
- Galbusera M, Remuzzi G, Boccardo P. Treatment of bleeding in dialysis patients. *Semin Dial* 2009;22:279-86.
- Goldenberg NA, Manco-Johnson MJ. Pediatric hemostasis and use of plasma components. *Best Pract Res Clin Haematol* 2006;19:143-55.
- Goldstein JN, Refaai MA, Milling TJ Jr, et al. Four-factor pro-thrombin complex concentrate versus plasma for rapid vitamin K antagonist reversal in patients needing urgent surgical or invasive interventions: A phase 3b, open-label, non-inferiority, randomised trial. *Lancet* 2015;385:2077-87.
- Goodnough LT, Shander A. How we treat management of warfarin-associated coagulopathy in patients with intracerebral hemorrhage. *Blood* 2011;117:6091-9.
- Gosselin RC, Marshall C, Dwyre DM, et al. Coagulation profile of liquid-state plasma. *Transfusion* 2013;53:579-90.
- Gulati G, Hevelow M, George M, et al. International normalized ratio versus plasma levels of coagulation factors in patients on vitamin K antagonist therapy. *Arch Pathol Lab Med* 2011;135(4):490-4.
- Hadjesfandiari N, Levin E, Serrano K, et al. Risk analysis of transfusion of cryoprecipitate without consideration of ABO group. *Transfusion* 2021;61(1):29-34.
- Hardy J-F, de Moerloose P, Samama CM. Massive transfusion and coagulopathy: Pathophysiology and implications of clinical management. *Can J Anesth* 2006;53:S40-58.
- Heim KF, Flesher TA, Stroneck DF, et al. The relationship between alloimmunization and posttransfusion granulocyte survival: Experience in a chronic granulomatous disease cohort. *Transfusion* 2011;51:1154-62.

- Holland LL, Brooks JP. Toward rational fresh frozen plasma transfusion: The effect of plasma transfusion on coagulation test results. *Am J Clin Pathol* 2006;126:133-9.
- Inbal A, Oldenburg J, Carcao M, et al. Recombinant factor XIII: A safe and novel treatment for congenital factor XIII deficiency. *Blood* 2012;119:5111-17.
- Ketchum L, Hess JR, Hiippala S. Indications for early fresh frozen plasma, cryoprecipitate, and platelet transfusion in trauma. *J Trauma* 2006;60(6 Suppl):S51-8.
- Khan J, Dunbar NM. Time to stop worrying about ABO incompatible cryoprecipitate transfusions in adults. *Transfusion* 2021;61(1):1-4.
- Kor DJ, Stubbs JR, Gajic O. Perioperative coagulation management—fresh frozen plasma. *Best Pract Res Clin Anaesthesiol* 2010;24:51-64.
- Lee SH, Lee SM, Kim CS, et al. Fibrinogen recovery and changes in fibrin-based clot firmness after cryoprecipitate administration in patients undergoing aortic surgery involving deep hypothermic circulatory arrest. *Transfusion* 2014;54:1379-87.
- Matijevic N, Wang Y-W, Cotton B, et al. Better hemostatic profiles of never-frozen liquid plasma compared with thawed fresh frozen plasma. *J Trauma Acute Care Surg* 2013;74:84-90.
- McCullagh J, Cardigan R, Brunskill SJ, et al. Assessing the risks of haemolysis as an adverse reaction following the transfusion of ABO incompatible plasma-containing components—A scoping review. *Blood Rev* 2022;56:100989.
- Medical and Scientific Advisory Council of the National Hemophilia Foundation. MASAC recommendations concerning products licensed for the treatment of hemophilia and other bleeding disorders. MASAC recommendation #249, May 19, 2017. New York: National Hemophilia Foundation, 2017.
- Neisser-Svae A, Trawnicek L, Heger A, et al. Five-day stability of thawed plasma: Solvent/detergent-treated plasma comparable with fresh-frozen plasma and plasma frozen within 24 hours. *Transfusion* 2016;56:404-9.
- Nichols WL, Hultin MB, James AH, et al. von Willebrand disease (VWD): Evidence-based diagnosis, and management guidelines, the NHLBI Expert Panel report. *Haemophilia* 2008;14:171-232.
- Norda R, Andersson T, Edgren G, et al. The impact of plasma preparations and their storage time on short-term posttransfusion mortality: A population-based study using the Scandinavian Donation and Transfusion database. *J Trauma Acute Care Surg* 2012;72:954-60.
- O'Neill EM, Rowley J, Hansson-Wicher M, et al. Effect of 24-hour whole-blood storage on plasma clotting factors. *Transfusion* 1999;39:488-91.
- O'Shaughnessy DF, Atterbury C, Bolton-Maggs P, et al for the British Committee for Standards in Haematology, Blood Transfusion Task Force. Guidelines for the use of fresh-frozen plasma, cryoprecipitate and cryosupernatant. *Br J Haematol* 2004;126:11-28. Amendments 2006;136:514-16.
- Pagano B, Morton S, Cohn C, et al. An international registry of granulocyte transfusions. *Transfus Med Hemother* 2018;45:318-22.
- Pearse BL, Smith I, Faulke D, et al. Protocol guided bleeding management improves cardiac surgery patient outcomes. *Vox Sang* 2015;109:267-79.
- Poterjoy BS, Josephson CD. Platelets, frozen plasma, and cryoprecipitate: What is the clinical evidence for their use in the neonatal intensive care unit? *Semin Perinatol* 2009;33:66-74.
- Price TH. Granulocyte transfusion: Current status. *Semin Hematol* 2007;44:15-23.
- Price TH, Boeckh M, Harrison RW, et al. Efficacy of transfusion with granulocytes from G-CSF/dexamethasone-treated donors in neutropenic patients with infection. *Blood* 2015;126:2153-61.
- Ramsey G. Treating coagulopathy in liver disease with plasma transfusions or recombinant factor VIIa: An evidence-based review. *Best Pract Res Clin Haematol* 2006;19:113-26.
- Raycraft T, Bartoszko J, Karkouti K, et al. Practice patterns of ABO-matching for cryoprecipitate and patient outcomes after ABO-compatible versus incompatible cryoprecipitate. *Vox Sang* 2022;117(9):1105-11.
- Roback JD, Caldwell S, Carson J, et al. Evidence-based practice guidelines for plasma transfusion. *Transfusion* 2010;50:1227-39.
- Rourke C, Curry N, Khan S, et al. Fibrinogen levels during trauma hemorrhage, response to replacement therapy, and association with patient outcomes. *J Thromb Hemost* 2012;10:1342-51.
- Scott EA, Puca KE, Bradley C, et al. Comparison and stability of ADAMTS13 activity in therapeutic plasma products. *Transfusion* 2007;47:120-5.
- Scott EA, Puca K, Heraly JC, et al. Evaluation and comparison of coagulation factor activity in fresh-frozen plasma and 24-hour plasma at thaw and after 120 hours of 1 to 6 C storage. *Transfusion* 2009;49:1584-91.

- Segal JB, Dzik WH. Paucity of studies to support that abnormal coagulation test results predict bleeding in the setting of invasive procedures: An evidence-based review. *Transfusion* 2005;45:1413-25.
- Spence RK. Clinical use of plasma and plasma fractions. *Best Pract Res Clin Haematol* 2006;19:83-96.
- Stanworth SJ, Brunskill SJ, Hyde CJ, et al. Is fresh frozen plasma clinically effective? A systematic review of randomized controlled trials. *Br J Haematol* 2004;126:139-52.
- Szumowski A, Radziwon PM. Granulocyte transfusion vs. neutropenia - is there a chance to win this battle? *Med Res J* 2022;7(4):321-32.
- Tanheco YC. Granulocyte transfusion therapy. *Ann Blood* 2022;7:5.
- Triulzi DJ. The art of plasma transfusion therapy. *Transfusion* 2006;46:1268-70.
- Triulzi D, Gottschall J, Murphy E, et al. A multicenter study of plasma use in the United States. *Transfusion* 2015;55:1313-19.
- Wehrli G, Taylor Ne, Haines AL, et al. Blood components: Instituting a thawed plasma procedure: It just makes sense and saves cents. *Transfusion* 2009;49:2625-30.
- Yazer MH, Cortese-Hassett A, Triulzi DJ. Coagulation factor levels in plasma frozen within 24 hours of phlebotomy over 5 days of storage at 1 to 60 C. *Transfusion* 2008;48:2525-30.

Platelets

- Ashford P, Gulliksson H, Georgsen J, Distler P. Standard terminology for platelet additive solutions. *Vox Sang* 2010;98:577-8.
- Blumberg N, Heal JM, Rowe JM. A randomized trial of washed red blood cell and platelet transfusions in adult leukemia. *BMC Blood Disord* 2004;4:6.
- Brecher M. The platelet prophylactic trigger: When expectations meet reality. *Transfusion* 2007;47:188-91.
- British Committee for Standards in Haematology, Blood Transfusion Task Force. Guidelines for the use of platelet transfusions. *Br J Haematol* 2003;122:10-23.
- British Committee for Standards in Haematology, Blood Transfusion Task Force. Transfusion guidelines for neonates and older children. *Br J Haematol* 2004;124:433-53.
- Cid J, Lozano M, Ziman A, et al. Biomedical Excellence for Safer Transfusion collaborative (2015). Low frequency of anti-D alloimmunization following D+ platelet transfusion: The Anti-D Alloimmunization after D-incompatible Platelet Transfusions (ADAPT) study. *Br J Haematol* 2015;168(4):598-603.
- Estcourt LJ, Malouf R, Murphy MF. Pathogen-reduced platelets for the prevention of bleeding in people of any age. *JAMA Oncol* 2018;4:57.
- Gottschall J, Wu Y, Triulzi D, et al; NHLBI Recipient Epidemiology and Donor Evaluation (REDS-III) Study. The epidemiology of platelet transfusions: An analysis of platelet use at 12 US hospitals. *Transfusion* 2020; 60(1):46-53.
- Heal JM, Blumberg N. Optimizing platelet transfusion therapy. *Blood Rev* 2004;31:1-14.
- Herve F, Tardivel R, Semana G, et al. Large scale use of platelet additive solution (PAS) reduces allergic type transfusion adverse events. *Vox Sang* 2007;93(Suppl 1):267.
- Kerkhoffs JL, Eikenboom JC, Schipperus MS, et al. A multicenter randomized study of the efficacy of transfusion with platelets stored in platelet additive solution II versus plasma. *Blood* 2006;108:3210-15.
- Kerkhoffs JL, van Putten WL, Novotny VM, et al. Clinical effectiveness of leucoreduced, pooled donor platelet concentrates, stored in plasma or additive solution with and without pathogen reduction. *Br J Haematol* 2010; 150:209-17.
- McCullough J. Overview of platelet transfusion. *Semin Hematol* 2010;47:235-42.
- O'Brien KL, Haspel RL, Uhl L. Anti-D alloimmunization after D-incompatible platelet transfusions: A 14-year single-institution retrospective review. *Transfusion* 2014;54(3):650-4.
- Schiffer CA, Bohlke K, Delaney M, et al. Platelet transfusion for patients with cancer: American Society of Clinical Oncology clinical practice guideline update. *J Clin Oncol* 2018;36(3):283-99.
- Schoenfeld H, Muhm M, Doepfner U, et al. The functional integrity of platelets in volume-reduced platelet concentrates. *Anesth Analg* 2005;100:78-81.
- Schoenfeld H, Spies C, Jakob C. Volume-reduced platelet concentrates. *Curr Hematol Rep* 2006;5:82-8.

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